Original Article

Expression level of plasma Bcl-xL and Bcl-2 in patients with systemic lupus erythematosus

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Received November 19, 2015; Accepted February 3, 2016; Epub March 15, 2016; Published March 30, 2016

Abstract: Objective: To investigate the plasma levels of Bcl-xL and Bcl-2 in systemic lupus erythematosus (SLE) patients and their correlations with SLE-associated clinical parameters. Methods: SLE patients and 80 normal controls were recruited, the plasma of them were obtained at the first visit. Plasma Bcl-xL and Bcl-2 levels were evaluated by Enzyme-Linked Immunosorbent Assay (ELISA). Results: When compared with the normal controls, the plasma level of Bcl-xL was significantly lower in SLE patients, while the plasma level of Bcl-2 in SLE patients was significantly higher (both P<0.05). Furthermore, the plasma level of Bcl-xL was positively associated with the level of C4 and was inversely correlated with the levels of IgG and IgM. Conclusion: All these findings suggest Bcl-xL and Bcl-2 possible role in the pathogenesis of SLE. However, further studies are needed to confirm these preliminary results.

Keywords: Bcl-xL, Bcl-2, systemic lupus erythematosus

Introduction

Systemic lupus erythematosus (SLE) is a chronic, multifactorial autoimmune disease with the features of the presence of multiple autoantibodies, complement activation, and immunecomplex deposition, causing serious injury to various organs or systems [1-3]. So far, although the exact cause of SLE remains unclear, it is sure that the disorder of B cell and T cell immune function plays a key role for the disease pathogenesis, especially B cell.

Bcl-xL and Bcl-2 are inhibitors of apoptotic cell death belonging to the Bcl-2 family proteins. Dysregulated expression of either of the two molecules may result in autoimmunity. Transgenic expression of Bcl-xL or Bcl-2 in murine B cells was shown to modify the repertoire of B cells and to result in the production of pathogenic antibodies and the development of autoimmune diseases including SLE [4, 5]. But The Bcl-xL and Bcl-2 transgenes also act differently during negative selection in immature B cells, as transgenic Bcl-xL has the ability to block

negative selection and promote developmental maturation, whereas autoreactive cells transgenic for Bcl-2 remain arrested in development [4, 6]. Fang et al demonstrated that the Bcl-xL transgene allowed self-reactive B cells that normally arrest in development and die in the bone marrow (BM) to escape clonal deletion and proposed that Bcl-xL may have a distinct role in controlling survival at the immature stage of B cell development by generating B cell-restricted Bcl-xL transgenic mice [4]. Bcl-xL also play a role in the development, differentiation, and clonal selection of B cells [7, 8]. Mice expressing a Bcl-2 transgene in their B-cells were prone to develop an autoimmune disease resembling SLE [5]. Together, all these evidences indicated that Bcl-xL and Bcl-2 may be involved in SLE pathogenesis. However, the association studies between Bcl-2 SNP markers and SLE have shown different results among different populations [9, 10]. And previous studies has indicated that the expression of Bcl-2 varies among various cells [11-14]. Therefore, the role of BclxL and Bcl-2 in SLE remain to be further elucidated.

Table 1. The general features of study subjects

Parameters	SLE patients (n=80)	Healthy control (n=80)	P value
Age	37.81 ± 14.50	35.48 ± 8.97	0.222
Sex (male/female)	7/73	11/69	0.317
Active (SLEDAI ≥10)	43/37	NA	NA
Nephritis (yes/no)	27/53	NA	NA

Note: SLEDAI: SLE disease activity index; NA: not applicable.

Table 2. Comparison of plasma BCL-XL levels and plasma BCL-2 levels between different subgroups

Number	Plasma level (ng/ ml) Mean ± SD	P value
74	74.70 ± 56.80	
51	32.79 ± 16.40	<0.001*
35	30.85 ± 16.95	
16	37.03 ± 14.75	0.215☆
23	34.15 ± 18.22	
28	31.67 ± 15.00	0.596
71	56.38 ± 36.44	
73	68.56 ± 30.36	0.031*
49	70.69 ± 33.23	
24	64.21 ± 23.48	0.341☆
33	73.27 ± 37.32	
40	64.67 ± 22.90	0.253
	74 51 35 16 23 28 71 73 49 24 33	74 74.70 ± 56.80 51 32.79 ± 16.40 35 30.85 ± 16.95 16 37.03 ± 14.75 23 34.15 ± 18.22 28 31.67 ± 15.00 71 56.38 ± 36.44 73 68.56 ± 30.36 49 70.69 ± 33.23 24 64.21 ± 23.48 33 73.27 ± 37.32

Note: *versus normal controls; *versus SLE without nephritis; $^{\blacktriangle} \text{versus}$ less active SLE.

In the present study, to further explore the role of Bcl-xL and Bcl-2 in human SLE, we investigated the Bcl-xL and Bcl-2 plasma levels in SLE and their relations with SLE-associated clinical parameters.

Materials and methods

Patients

SLE patients were recruited from the Department of Rheumatology, First Affiliated Hospital of Anhui Medical University and the Department of Rheumatology, Anhui Provincial Hospital. All the patients fulfilled the requirement of at least four criteria of American College of Rheumatology (ACR) classification for SLE [15]. Individual disease activity was scored by the SLE Disease Activity Index (SLEDAI) [16]. The patients with renal disease were defined by persistent proteinuria (0.5

g/24 h) or the presence of active cellular casts; or biopsy evidence of lupus nephritis [17]. Demographic, clinical, and laboratory data were collected from hospital records or by questionnaire and reviewed by experienced physicians. All of 80 controls were clinically assessed to be without SLE, other autoimmune disorders, systemic disorders or family history of autoimmune disorders (including first-, second- and third-degree relatives). Informed consent for the studies was provided by all participants in accordance with the protocol as approved by the Ethics Committee of each hospital.

Method

Extraction of serum and enzyme-linked immunosorbent assay (ELISA): Serum were obtained from 5 ml of whole blood at the first visit of the study subjects and stored at -80°C. Plasma levels of Bcl-xL and Bcl-2 were determined using commercially available sandwich enzyme linked immunosorbent assay (ELISA) developing kits. All kits were derived from R&D Systems (Ambingdon, UK). The assay procedure was performed according to the manufacturer's instructions of the enclosed pamphlet.

Statistical analysis

The numerical data were expressed as mean \pm SD. For comparing the means between two different groups, the Student's t-test was used. All statistical analysis was conducted by the Statistical Package for the Social Sciences (SPSS) statistical software for Windows, Version 10.01 (SPSS Inc, IL, USA). P Values less than 0.05 were considered statistically significant.

Results

Characteristics of participants

The mean ages of the SLE subjects were 37.81 \pm 14.50 years, while the mean ages of controls was 35.48 \pm 8.97 years. 73 SLE patients (91%) and 69 subjects (86%) in control group were female. There were no significant differences between patient group with SLE and healthy controls in terms of age and gender distribution (P = 0.222, P = 0.317, respectively) (**Table 1**).

Table 3. Associations of plasma BCL-XL and BCL-2 levels with clinical parameters of SLE patients

Group	-/+	Number	Plasma level (pg/ ml) Mean ± SD	P value
BCL-XL				
Vasculitis	-	46	32.63 ± 16.49	0.842
	+	5	34.19 ± 17.44	
Arthritis	-	34	34.45 ± 17.19	0.311
	+	17	29.26 ± 14.62	
Myositis	-	44	33.57 ± 16.89	0.397
	+	7	27.85 ± 12.85	
Rash	-	24	31.53 ± 19.21	0.610
	+	27	33.91 ± 13.73	
Alopecia	-	30	29.74 ± 16.79	0.114
	+	21	37.13 ± 15.18	
Oral ulcer	-	41	34.63 ± 15.63	0.104
	+	10	25.22 ± 18.18	
Pericarditis	-	47	32.68 ± 16.45	0.876
	+	4	34.04 ± 18.25	
Fever	-	37	32.31 ± 16.93	0.738
	+	14	34.05 ± 15.47	
BCL-2				
Vasculitis	-	68	68.08 ± 31.14	0.624
	+	5	75.04 ± 16.85	
Arthritis	-	49	68.99 ± 32.98	0.849
	+	24	67.67 ± 24.77	
Myositis	-	66	67.56 ± 31.19	0.393
	+	7	77.96 ± 20.22	
Rash	-	36	75.09 ± 33.90	0.069
	+	37	62.20 ± 25.31	
Alopecia	-	45	70.50 ± 30.36	0.494
	+	28	65.45 ± 30.65	
Oral ulcer	-	60	68.51 ± 32.09	0.977
	+	13	38.78 ± 21.63	
Pericarditis	-	68	68.74 ± 31.29	0.727
	+	5	66.16 ± 13.46	
Fever	-	53	71.03 ± 31.62	0.261
	-	20	62.02 ± 26.36	

Note: +/-: with/without.

The plasma levels of Bcl-xL and Bcl-2 in SLE patients and controls

The plasma level of Bcl-xL was lower in SLE patients than in normal controls (32.79 \pm 16.40 vs. 74.70 \pm 56.80, P<0.001). Compared with the normal controls, SLE patients had a higher level of Bcl-2 (68.56 \pm 30.36 vs. 56.38 \pm 36.44, P = 0.031) (**Table 2**).

Association of Bcl-xL and Bcl-2 plasma levels with SLE-associated clinical parameters

Both plasma levels of Bcl-xL and Bcl-2 had no significant correlations with LN, SLEDAI score, SLE-associated manifestations including arthritis, vasculitis, rash, alopecia, oral ulcer, pericarditis and fever (Table 3), categorical laboratory parameters including Anti-dsDNA, Anti-Sm, Anti-SSA, Anti-SSB, Anti-RNP, Anti-RibosomalP, thrombocytopenia, leukopenia, proteinuria, and blood urine (Table 4) and some quantitative laboratory parameters including C3, C-reactive protein(CRP), and IgA. The plasma level of Bcl-xL was correlated with the levels of C4 (r = 0.293, P = 0.041) and was negatively correlated with the levels of IgG (r = -0.340, P = 0.024), IgM (r = -0.352, P = 0.019) (Table 5).

Discussion

In recent years, there has been increasing studies on the role of proteins of the Bcl-2 family in regulating apoptotic cell death. Bcl-xL and Bcl-2 are two numbers of the Bcl-2 family proteins participating in the regulating B cell immune response. During B cell ontogeny, Bcl-xL is expressed at highest levels in the small pre-B cell stage of development [18], but at very low levels in pro-B cells and downregulated in immature and mature B resting cells [18, 19]. Fang et al found that the death of pro-B cells with failed immunoglobulin rearrangements occurred by apoptosis and large expansions of pro-B cells developed in bone marrow in transgenic mice expressing Bcl-xL gene in the B lineage. And Bcl-xL can deliver a strong survival signal at the pro-B stage [4]. Takahashi generated Bcl-xL transgene mice in which endogenous Bcl-xL was produced [7]. The transgene mice demonstrated reduced apoptosis in germinal centers B cells and resulted in the expansion of B lymphocytes bearing VDJ rearrangements. The abundance of these noncanonical cells lowered the average affinity of long-lived antibody-forming cells and serum antibody, demonstrating that Bcl-xL influence clonal selection/maintenance for affinity maturation. All these results indicate that Bcl-xL may plays an important role in extending B cells survival, and also reveal an unexpected role of the BclxL in the regulation of B cells function and maintaining self-tolerance. Bcl-2 is highly ex-

Table 4. Associations of plasma BCL-XL and BCL-2 levels with categorical laboratory parameters of SLE patients

BCL-XL Anti-dsDNA - 27 32.82 ± 17.21 0.988 + 24 32.75 ± 15.82 Anti-Sm - 36 35.01 ± 16.05 0.134 + 15 27.44 ± 16.54 Anti-SSA - 14 32.87 ± 21.50 0.985 + 37 32.75 ± 14.38 Anti-SSB - 42 75.04 ± 16.85 0.539 + 9 32.77 ± 21.10 Anti-RNP - 32 34.23 ± 15.17 0.419
Anti-dsDNA - 27 32.82 ± 17.21 0.988 + 24 32.75 ± 15.82 Anti-Sm - 36 35.01 ± 16.05 0.134 + 15 27.44 ± 16.54 Anti-SSA - 14 32.87 ± 21.50 0.985 + 37 32.75 ± 14.38 Anti-SSB - 42 75.04 ± 16.85 0.539 + 9 32.77 ± 21.10
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Anti-Sm - 36 35.01 ± 16.05 0.134 + 15 27.44 ± 16.54 Anti-SSA - 14 32.87 ± 21.50 0.985 + 37 32.75 ± 14.38 Anti-SSB - 42 75.04 ± 16.85 0.539 + 9 32.77 ± 21.10
+ 15 27.44 ± 16.54 Anti-SSA - 14 32.87 ± 21.50 0.985 + 37 32.75 ± 14.38 Anti-SSB - 42 75.04 ± 16.85 0.539 + 9 32.77 ± 21.10
Anti-SSA - 14 32.87 ± 21.50 0.985 + 37 32.75 ± 14.38 Anti-SSB - 42 75.04 ± 16.85 0.539 + 9 32.77 ± 21.10
+ 37 32.75 ± 14.38 Anti-SSB - 42 75.04 ± 16.85 0.539 + 9 32.77 ± 21.10
Anti-SSB - 42 75.04 ± 16.85 0.539 + 9 32.77 ± 21.10
+ 9 32.77 ± 21.10
Anti-RNP - 32 34.23 ± 15.17 0.419
+ 19 30.35 ± 18.47
Anti-RibosomalP - 33 34.18 ± 16.03 0.418
+ 18 30.24 ± 17.24
Thrombocytopenia - 36 31.15 ± 14.49 0.275
+ 15 36.70 ± 20.32
Leukopenia - 40 32.41 ± 15.68 0.759
+ 11 34.15 ± 19.61
Proteinuria - 32 31.41 ± 17.24 0.444
+ 19 35.10 ± 15.06
Blood urine - 36 31.06 ± 17.09 0.248
+ 15 36.93 ± 14.32
BCL-2
Anti-dsDNA - 40 72.09 ± 33.66 0.277
+ 33 64.28 ± 25.65
Anti-Sm - 53 67.14 ± 31.48 0.518
+ 20 72.33 ± 27.56
Anti-SSA - 22 70.65 ± 35.78 0.703
+ 51 37.66 ± 28.04
Anti-SSB - 61 68.61 ± 31.56 0.776
+ 12 38.30 ± 24.46
Anti-RNP - 44 38.24 ± 29.99 0.913
+ 29 39.04 ± 31.43
Anti-Ribosomal P - 52 67.14 ± 31.63 0.534
+ 21 72.07 ± 27.34
Thrombocytopenia 56 66.14 ± 29.62 0.219
17 76.52 ± 32.30
Leukopenia 59 66.07 ± 31.09 0.152
14 79.05 ± 25.39
Proteinuria 47 70.79 ± 31.53 0.402
26 64.52 ± 28.26
Blood urine - 51 72.37 ± 32.85 0.057
- 22 59.72 ± 21.76

Note: +/-: with/without.

pressed in the pro-B cells and mature B lymphocytes, but downregulated at the pre-B and

immature B cell stages of maturation [20, 21]. In a previous study, Strasser et al generated a transgenic mice harboring human Bcl-2 cDNA under the control of an immunoglobulin heavy chain enhancer (Eµ) which was replaced by a representative transgenic strain Eu-bcl-2-22 [22]. The mutant mice enforced Bcl-2 expression demonstrated a great excess of B lymphocytes, immunoglobulinsecreting cells, and serum inmmunoglobulins, attributable to increased longevity of B-lineage cells. Pre-B and plasma cells as well as B cells exhibited prolonged survival in culture in the mutant mice. Thus ES-Bcl-2-22 mice constitute a transgenic model for a systemic autoimmune disease resembling human SLE. Collectively, Bcl-xL and Bcl-2 may play an important role in the induction and development of autoimmune diseases.

SLE is considered to be the prototype of human autoimmune diseases. It is a disorder of generalized autoimmunity characterized by multisystem organ involvement, polyclonal B cell activation, and the production of autoantibodies against nuclear, cytoplasmic, and cell surface antigens [10]. More recently. Zhan et al. investigated the effects of two inhibitory anti-Bcl-2 (ABT-737, ABT-199) in lupusprone NZB/W F1 mice and human peripheral blood including SLE and controls in vitro, their results showed that ABT-199 treatment efficiently killed NZB/W plasmacytoid DCs (pDCs) which are major producers of IFN-α and also being a prominent source of B cell activating factor of the tumor necrosis factor family (BAFF) [23]. As we known, IFN- α and BAFF had been proposed to contribute to SLE pathogenesis [24, 25]. Thus they proposed that enhanced cytokine output from pDCs due to their extended survival, mediated by increased BCL-2 levels, could be a significant driver of SLE. All these results support a possible role of Bcl-xL and Bcl-2 in the pathogenesis of SLE.

In the current study, we investigated the plasma Bcl-xL and Bcl-2 levels in SLE patients. Results showed that lower level of Bcl-xL and higher level of Bcl-2 were observed

Table 5. Associations of plasma BCL-XL and BCL-2 levels with quantitative laboratory parameters of SLE patients

Parameters	N	Spearman Correlation coefficient (r/r_s)	P value
BCL-XL			
C3	80	0.216	0.128
C4	75	0.293	0.041
CRP	66	0.205	0.183
IgA	71	-0.289	0.057
IgG	71	-0.340	0.024
IgM	71	-0.352	0.019
SLEDAI	79	0.020	0.891
BCL-2			
C3	85	0.205	0.082
C4	79	-0.051	0.675
CRP	70	-0.226	0.082
IgA	75	0.177	0.160
IgG	75	-0.102	0.421
IgM	75	0.027	0.832
SLEDAI	84	-0.129	0.281

in the plasma of SLE patients compared with health controls. In addition, the plasma level of Bcl-xL was correlated with the levels of C4 and was negatively correlated with the levels of IgG, IgM in SLE patients. It is of interest to compare our results to several previous attempts to explore the expression and clinical associations of Bcl-xL and Bcl-2 in SLE patients. A previous study by Rose et al suggest that Bcl-2 expression is unaltered in unfractionated peripheral blood mononuclear cells in patients with SLE and Bcl-2 levels did not correlate with overall disease activity in SLE patients, which are similar to our results [26]. However, another study by Liphaus et al reported upregulation of Bcl-2 protein expression in peripheral blood mononuclear cells from patients with juvenileonset SLE, and the higher Bcl-2 expression, the active disease [11]. In addition, a decreased Bcl-2 expression in peripheral blood lymphocytes of patients with SLE have also been shown and Bcl was negatively correlated with disease activity [14]. These disparities may be influenced by the difference among various lymphocyte subsets, various disease subgroups, various cells, different demographic characteristics, application of different classification and patient selection criteria.

Taken together, decreased plasma level of Bcl-xL and increased plasma level of Bcl-2, and

association of Bcl-xL with C4, IgG and IgM suggest their possible role in the pathogenesis of SLE. However, further studies are needed to confirm these preliminary results.

Acknowledgements

This work was supported by grants from the Natural Science Foundation of Anhui Province (1408085QH179), and the National Natural Science Foundation of China (81573222, 81-473058).

Disclosure of conflict of interest

None.

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References

- [1] Cook HT and Botto M. Mechanisms of Disease: the complement system and the pathogenesis of systemic lupus erythematosus. Nat Clin Pract Rheumatol 2006; 2: 330-337.
- [2] Moulton VR and Tsokos GC. Abnormalities of T cell signaling in systemic lupus erythematosus. Arthritis Res Ther 2011; 13: 207.
- [3] Dorner T, Giesecke C and Lipsky PE. Mechanisms of B cell autoimmunity in SLE. Arthritis Res Ther 2011; 13: 243.
- [4] Fang W, Weintraub BC, Dunlap B, Garside P, Pape KA, Jenkins MK, Goodnow CC, Mueller DL and Behrens TW. Self-reactive B lymphocytes overexpressing Bcl-xL escape negative selection and are tolerized by clonal anergy and receptor editing. Immunity 1998; 9: 35-45.
- [5] Strasser A, Whittingham S, Vaux DL, Bath ML, Adams JM, Cory S and Harris AW. Enforced BCL2 expression in B-lymphoid cells prolongs antibody responses and elicits autoimmune disease. Proc Natl Acad Sci U S A 1991; 88: 8661-8665.
- [6] Hartley SB, Cooke MP, Fulcher DA, Harris AW, Cory S, Basten A and Goodnow CC. Elimination of self-reactive B lymphocytes proceeds in two stages: arrested development and cell death. Cell 1993; 72: 325-335.
- [7] Takahashi Y, Cerasoli DM, Dal Porto JM, Shimoda M, Freund R, Fang W, Telander DG, Malvey EN, Mueller DL, Behrens TW and Kelsoe G. Relaxed negative selection in germi-

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- nal centers and impaired affinity maturation in bcl-xL transgenic mice. J Exp Med 1999; 190: 399-410.
- [8] Amanna IJ, Dingwall JP and Hayes CE. Enforced bcl-xL gene expression restored splenic B lymphocyte development in BAFF-R mutant mice. J Immunol 2003; 170: 4593-4600.
- [9] Johansson C, Castillejo-Lopez C, Johanneson B, Svenungsson E, Gunnarsson I, Frostegard J, Sturfelt G, Truedsson L, Lofstrom B, Alcocer-Varela J, Lundberg I, Gyllensten UB, Alarcon-Segovia D and Alarcon-Riquelme ME. Association analysis with microsatellite and SNP markers does not support the involvement of BCL-2 in systemic lupus erythematosus in Mexican and Swedish patients and their families. Genes Immun 2000; 1: 380-385.
- [10] Mehrian R, Quismorio FP Jr, Strassmann G, Stimmler MM, Horwitz DA, Kitridou RC, Gauderman WJ, Morrison J, Brautbar C and Jacob CO. Synergistic effect between IL-10 and bcl-2 genotypes in determining susceptibility to systemic lupus erythematosus. Arthritis Rheum 1998; 41: 596-602.
- [11] Liphaus BL, Kiss MH, Carrasco S and Goldenstein-Schainberg C. Increased Fas and Bcl-2 expression on peripheral mononuclear cells from patients with active juvenile-onset systemic lupus erythematosus. J Rheumatol 2007; 34: 1580-1584.
- [12] Aringer M, Wintersberger W, Steiner CW, Kiener H, Presterl E, Jaeger U, Smolen JS and Graninger WB. High levels of bcl-2 protein in circulating T lymphocytes, but not B lymphocytes, of patients with systemic lupus erythematosus. Arthritis Rheum 1994; 37: 1423-1430.
- [13] Falcini F, Azzari C, Gelli VA, Luchetti M, Gabrielli A, Calzolari A, Pignone A, Generini S and Matucci Cerinic M. Reduction of bcl-2 in T cells during immunosuppressive therapy in patients with severe juvenile onset systemic lupus erythematosus. Clin Immunol 1999; 93: 59-64.
- [14] Chan EY, Ko SC and Lau CS. Increased rate of apoptosis and decreased expression of bcl-2 protein in peripheral blood lymphocytes from patients with active systemic lupus erythematosus. Asian Pac J Allergy Immunol 1997; 15: 3-7.
- [15] Hochberg MC. Updating the American College of Rheumatology revised criteria for the classification of systemic lupus erythematosus. Arthritis Rheum 1997; 40: 1725.

- [16] Bombardier C, Gladman DD, Urowitz MB, Caron D and Chang CH. Derivation of the SLEDAI. A disease activity index for lupus patients. The Committee on Prognosis Studies in SLE. Arthritis Rheum 1992; 35: 630-640.
- [17] Tan EM, Cohen AS, Fries JF, Masi AT, McShane DJ, Rothfield NF, Schaller JG, Talal N and Winchester RJ. The 1982 revised criteria for the classification of systemic lupus erythematosus. Arthritis Rheum 1982; 25: 1271-1277.
- [18] Fang W, Mueller DL, Pennell CA, Rivard JJ, Li YS, Hardy RR, Schlissel MS and Behrens TW. Frequent aberrant immunoglobulin gene rearrangements in pro-B cells revealed by a bcl-xL transgene, Immunity 1996; 4: 291-299.
- [19] Grillot DA, Merino R, Pena JC, Fanslow WC, Finkelman FD, Thompson CB and Nunez G. bcl-x exhibits regulated expression during B cell development and activation and modulates lymphocyte survival in transgenic mice. J Exp Med 1996; 183: 381-391.
- [20] Merino R, Ding L, Veis DJ, Korsmeyer SJ and Nunez G. Developmental regulation of the Bcl-2 protein and susceptibility to cell death in B lymphocytes. EMBO J 1994; 13: 683-691.
- [21] Li YS, Hayakawa K and Hardy RR. The regulated expression of B lineage associated genes during B cell differentiation in bone marrow and fetal liver. J Exp Med 1993; 178: 951-960.
- [22] Strasser A, Harris AW and Cory S. bcl-2 transgene inhibits T cell death and perturbs thymic self-censorship. Cell 1991; 67: 889-899.
- [23] Zhan Y, Carrington EM, Ko HJ, Vikstrom IB, Oon S, Zhang JG, Vremec D, Brady JL, Bouillet P, Wu L, Huang DC, Wicks IP, Morand EF, Strasser A and Lew AM. Bcl-2 antagonists kill plasmacytoid dendritic cells from lupus-prone mice and dampen interferon-alpha production. Arthritis Rheumatol 2015; 67: 797-808.
- [24] Hagberg N and Ronnblom L. Systemic Lupus Erythematosus-A Disease with A Dysregulated Type I Interferon System. Scand J Immunol 2015; 82: 199-207.
- [25] Vincent FB, Morand EF, Schneider P and Mackay F. The BAFF/APRIL system in SLE pathogenesis. Nat Rev Rheumatol 2014; 10: 365-373.
- [26] Rose LM, Latchman DS and Isenberg DA. Bcl-2 expression is unaltered in unfractionated peripheral blood mononuclear cells in patients with systemic lupus erythematosus. Br J Rheumatol 1995; 34: 316-320.