

Original Article

Resveratrol inhibition of TNF- α and IL-1 for treatment of rheumatoid arthritis: from *In-Silico* to *In-vitro* elucidation

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Abstract: Resveratrol, a traditional Chinese medicine, known for its anti-inflammatory effect via cytokine inhibition, has long been tested and used in treating rheumatoid arthritis (RA). Here in this study we are exploring *In-silico* approaches to understand the mechanistic inhibition of TNF α and IL-1 cytokines using molecular docking and molecular simulation approach. The pathway analysis using GeneMania of these two cytokines showed us their importance in the network of protein involved in RA. The molecular docking showed mechanistic inhibition of TNF- α and IL-1 by resveratrol and showed us that TNF- α is a better suited target for resveratrol inhibition for RA treatment. The molecular dynamics simulations showed the complex of resveratrol and TNF- α to be stable over a run of five nano seconds in an environment close to *in-vivo* conditions. The study is corroborating resveratrol to be a potential RA drug. The IC50 of the resveratrol was determined to be 0.65 μ M.

Keywords: Resveratrol, rheumatoid arthritis, TNF- α , IL-1, docking, simulations, immunomodulator activity

Introduction

Rheumatoid arthritis (RA) an arthropathy of unknown etiology is a common inflammatory disease [1-4]. The etiology of the disease is governed by multiple factors, among smoking, obesity, drinking and genetic factors are well documented [5, 6]. The activated innate immune cells in RA patients release a spectrum of pro-inflammatory mediators: tumor necrosis factor alpha (TNF α), interleukin 1 (IL-1) [7-9], these mediators have a progressive role in joint inflammation and tissue destruction by inducing the production of metalloproteinases [3]. The concentration of these two cytokines is high in synovial fluid and the plasma in RA patients [10, 11].

Targeting these two cytokines has been attempted, TNF α inhibitors, anti-TNF antibody, a soluble TNF receptor fusion protein and IL-1 receptor antagonist have tried and tested [12, 13]. Compounds of natural origin have been used to target cytokines for treatment of RA [14] the compound of our interest here is 3,5,4'-trihydroxystilbene also known as resveratrol, a plant phytoalexin [15, 16]. Resveratrol

is known to be present in grape skin and red wine and has been reported to reduce superoxides, suppress carcinogenesis, angiogenesis, diabetes mellitus and inhibit inflammation by blocking TNF α and or IL-1 [17-20].

Computer aided drug designing (CADD) has become an important tool in drug discovery [21], and in this study we are using this approach to demonstrate the mechanistic inhibition of TNF α and IL-1 cytokines by resveratrol at atomic level. The molecular docking approach has been used for the atomic insight [22].

Material and methods

Protein preparation and pathway analysis

The TNF α and IL-1 cytokines proteins three dimensional (3D) structure were retrieved from PDB (<http://www.pdb.org/>). The TNF- α protein structure contains contained 147 amino acids ranging from 10-157, its PDB ID: is 2AZ5 [23]. IL-1 crystallographic structure has 154 amino acids ranging from 117-271, the PDB ID: 2ILA is assigned to it [24]. All the modifications prior to molecular docking were done using Discovery

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Figure 1. Protein sub networks of TNF- α and IL-1 (Query proteins) with IL-6, NFKB1A, MAP3K7, TGFB3 and TRAF6, some important proteins involved in RA. Only significant sub network is shown in the figure.

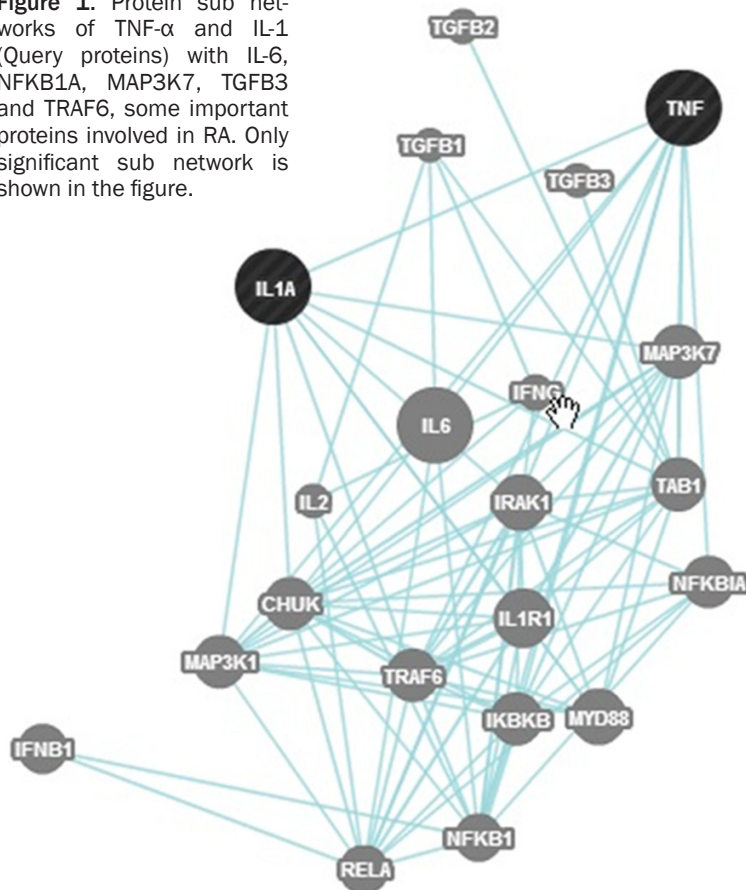


Table 1. The binding pattern of resveratrol with TNF- α and IL-1, number of interactions and distance calculated by pymol

Target Protein	ΔG (Kcal/mole)	No. of Interactions	Distance (Å)
TNF- α	-4.95	Four	Resveratrol-ASP45 (2.1Å)
			Resveratrol-GLN47 (3.0Å)
			Resveratrol-ILE136 (2.3Å)
			Resveratrol-ILE136 (3.0Å)
IL-1	-4.43	Four	Resveratrol-LEU40 (3.1Å)
			Resveratrol-ILE74 (2.8Å)
			Resveratrol-LEU79 (2.1Å)
			Resveratrol-VAL140 (2.3Å)

Studio 3.5 Visualizer [25]. The pathway of these two proteins was tracked using GENEMANIA server (www.genemania.org).

Compound selection

Resveratrol or 3,5,4'-trihydroxystilbene, a natural compound was retrieved from PubChem compound database. The SDF format of the compound was received bearing CID 445154.

The SDF format was converted to PDF for further study using Discovery Studio.

Molecular docking analysis

Molecular docking analysis of the TNF- α and IL-1 cytokine proteins with resveratrol was done. The top interactions out of ten were studied for each protein. Autodock 4.2 [26] tool was used for the study, the tool uses binding free energy evaluation to find the best binding mode between the compound and the protein. All the visualization were performed in Pymol [27].

Molecular dynamics simulation

The top compound selected was subjected to Molecular dynamics Simulations (MDS) using Gromacs 4.5.3 package [28]. Root mean square deviation (RMSD) root mean square fluctuation (RMSF) was checked using gromacs inbuilt tools `g_rms` and `g_rmsf` were used for the respective analysis.

Bioactivity

Immunomodulator activity of resveratrol was evaluated using oxidative burst assay. The standard protocol of chemiluminescence on reaction mixtures contained neutrophils was followed. Heparinized blood of healthy volunteers (18-45 years age) was used to purify neutrophils by dextran sedimentation and density gradient centrifugation. Neutrophils were adjusted to their required concentration using Hank's Balance Salt Solution containing Ca and Mg (HBSS++). 25 μ l of neutrophil cells were incubated with 25 μ l of serially diluted resveratrol with concentrations ranging from 0.1 μ M to 10 μ M. Control wells had neutrophil and HBSS++ but no resveratrol. Cells were washed after 30 min incubation with resveratrol HBSS++. Cells were activated by

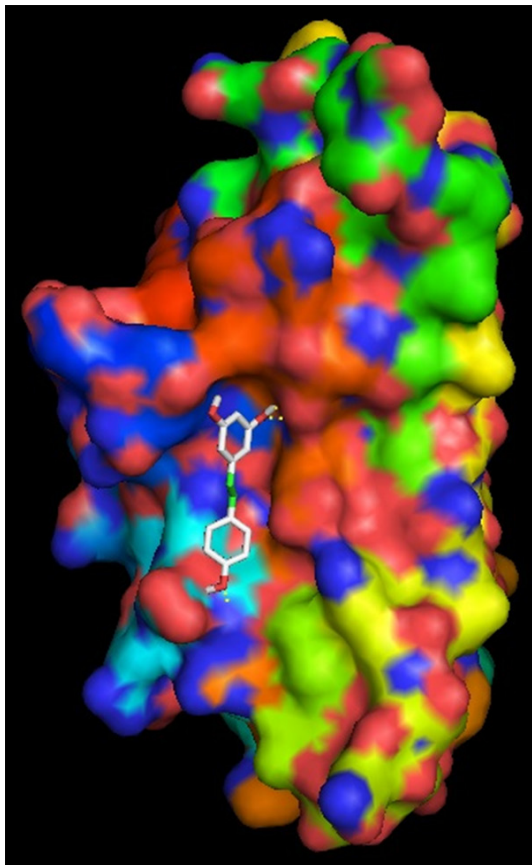


Figure 4. The best interaction of resveratrol with TNF- α , the complex used for MDS analysis.

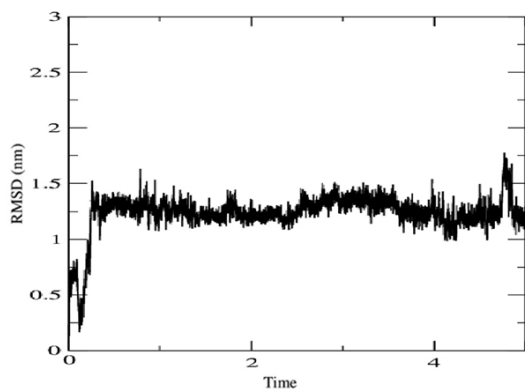


Figure 5. RMSD of the complex of TNF- α and resveratrol at constant temperature and pressure of 300 K and 1 bar. Showing the complex to be stable over a run of 5 ns.

Result

The pathway interaction profile of TNF- α and IL-1 derived using GeneMania database, web-based integrative software, gave us the net-

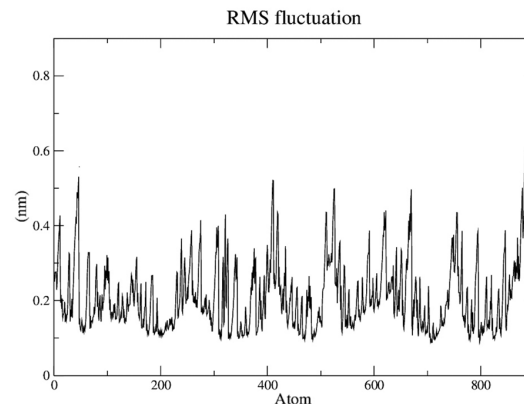


Figure 6. Atomic RMSF of the TNF- α in complex with resveratrol, showing the atomic fluctuation in the cytokine over a run of 5 ns.

work architecture of the connections of the two cytokines and other proteins of main importance, as shown in **Figure 1**. The modified crystallographic structures of TNF- α and IL-1 were subjected to molecular docking using AutoDock 4.2. The results generated were analyzed using Pymol, the measurement tool of the Pymol was also used to calculate the distances of the hydrogen bond, which are shown in **Table 1**. The first cytokine to be evaluated was TNF- α . The resveratrol and TNF- α complex, **Figure 2A** was selected from the ten complexes generated by the AutoDock suit based on the binding energy ΔG of -4.95 Kcal/mol. The complex is forming four hydrogen bonds with GLN47, ASP45 and two with ILE136 shown **Figure 3A**. The H₂₉ position of resveratrol is interacting with O of ASP45 and the bond formed between them has a distance of 2.1 Å. The N of GLN47 is forming a hydrogen bond of distance 3 Å with H at 27th position of resveratrol. The other two hydrogen bonds are formed by ILE136 with oxygen and H₂₇ of resveratrol, with a distance of 3 and 2.3 Å respectively. The second complex of resveratrol and IL-1, **Figure 2B** is the best of the top ten results generated with the ΔG of -4.43 Kcal/mol. This complex is also forming four hydrogen bonds with LEU40, ILE74, LEU79 and VAL140 as shown in **Figure 3B**, where LEU40, ILE74, LEU79 and VAL140 amino acids of IL-1 are forming hydrogen bond with O₃, H₂₇, H₂₉ and O₁ atoms of resveratrol respectively. The Hydrogen bond distance is 3.1 Å between LEU40 and O₃, 2.8 Å between ILE74 and H₂₇, 2.1 Å between LEU79 and H₂₉, 2.3 Å between VAL140 and O₁. This complex is the second

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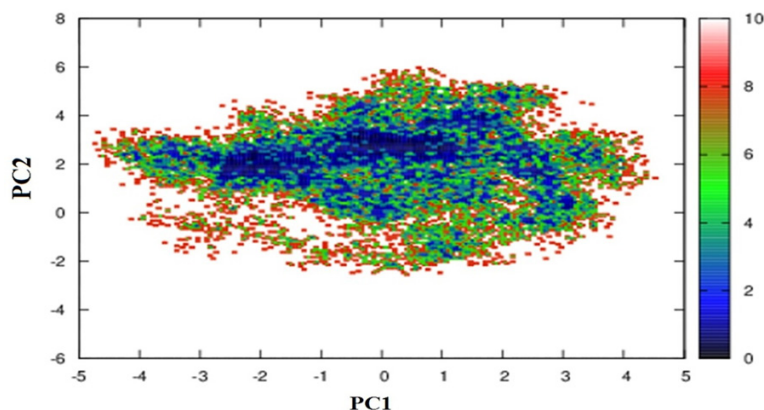


Figure 7. PC 1 vs. PC 2 of resveratrol-TNF complex.

Table 2. Immunomodulating inhibitory properties of resveratrol of neutrophil cells

Compound	Con. (μ M)	RLU mean \pm S.D.	Inh. %
Resveratrol	10	204.2 \pm 0.6	71.5
	1	313.5 \pm 2.1	64.1
	0.5	376.9 \pm 12.9	52.8
	0.15	580.1 \pm 12.2	22.5
	0.1	837.1 \pm 23.6	4.0
Control		872.3 \pm 83.6	

Con. = concentration. Inh.% = inhibition%.

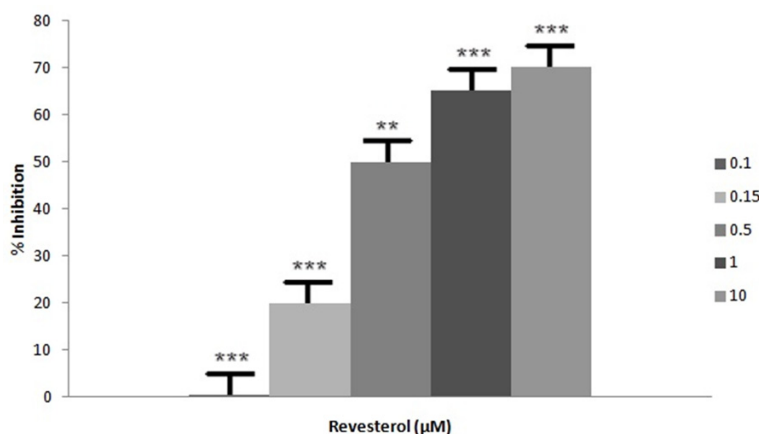


Figure 8. Eight concentration of the resveratrol (0.1 to 10 μ M) showing the inhibitory effect on oxidative burst as measured by chemiluminescence. The experiments were repeated five times.

best based on the binding energy. The top complex of TNF-resveratrol was subjected to MDS for further analysis. Five ns run was performed on this complex (Figure 4), prior to the run the complex was energy minimized for a period of 1 ns and a position restraints simulation was

performed using steepest decent integrator on a step-wise manner. The RMSD and RMSF of the complex was checked (Figure 5). The RMSD of the complex after 5 ns of MD simulation was showing a stable run at 1.25 \AA , showing very slight variation and concluding the complex to be stable throughout the run. The RMSF of the protein is represented by black in Figure 6 showing the fluctuation throughout the atoms of the protein. g-covar and g-anaeig of gromacs utilities was used to obtain the trajectories. In Figure 7 the projection PC 1 vs. PC 2 of the top complex is shown, their free energy surface was plotted the depiction shows that the stability of the complex over the run is uniform over time. To analyse the immunomodulator activity of resveratrol, the human neutrophils isolated from healthy individuals were incubated with several concentration of resveratrol for five minutes. The chemiluminescence was measured using luminol as probe (Table 2). Resveratrol possess average inhibitory effects in the range of 71.5, 64.1, 52.8, 22.5 and 4% on monocytes, respectively, at 10, 1, 0.5, 0.15 and 0.1 μ M concentration. Figure 8 shows % inhibition results to be dose dependent. The minimum concentration of resveratrol to show immunomodulation is 0.15 μ M and IC₅₀ of 0.65 μ M was determined.

Discussion

Natural products are said to be a gold mine for treatment for arthritis [14, 15] and our work explores resveratrol (3,4',5 trihydroxystilbene) a natural product of spermatophytes. The prod-

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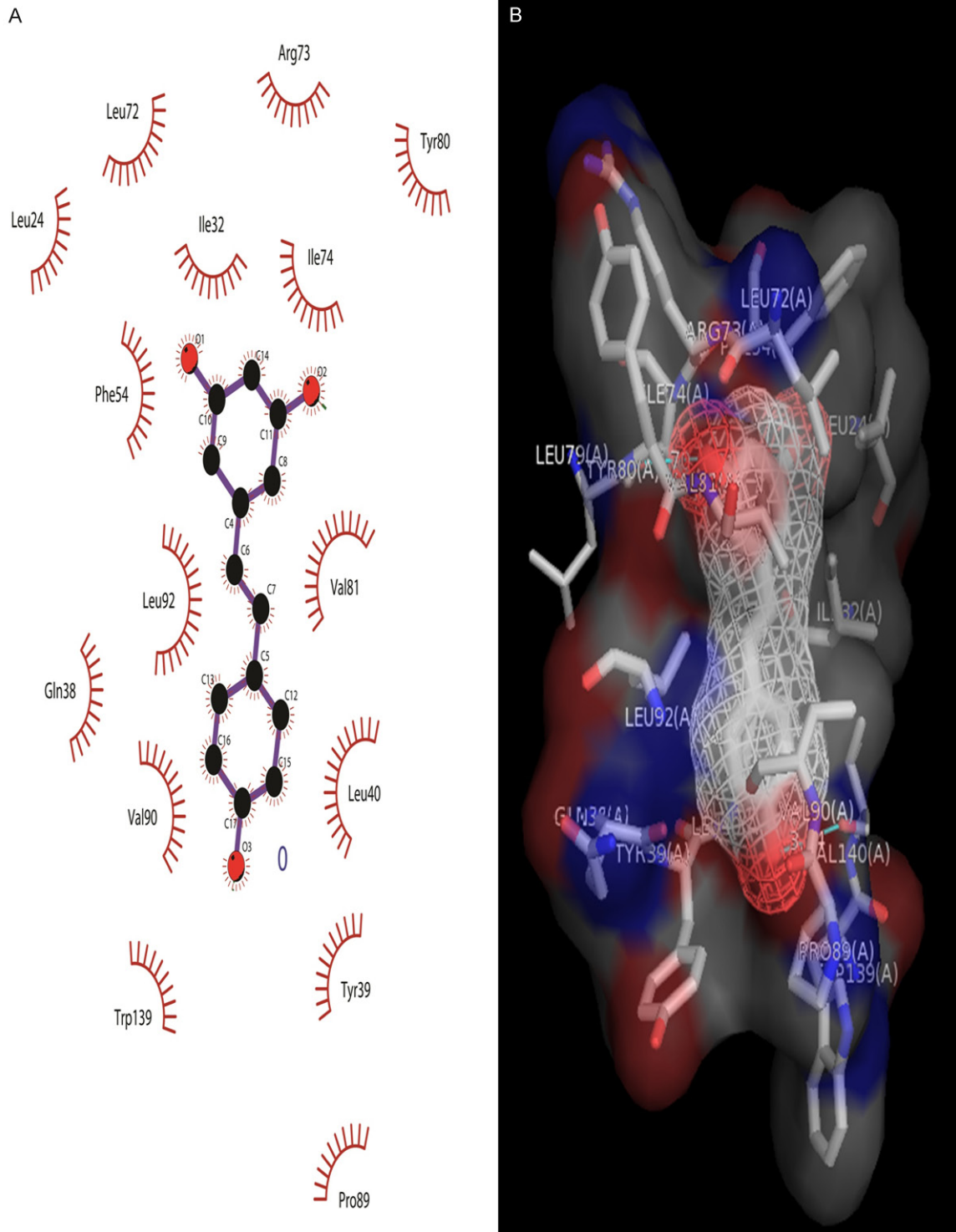


Figure 9. A. The hydrophobic interaction between the resveratrol and TNF- α protein target. B. The three dimensional representation of the same plot generated using Pymol.

uct has long been in use for treatment of RA and work explores its possible protein targets using advanced Insilico techniques. The impor-

tant proteins that are involved in the network and have a role in RA are IL-6, NFKB1A, MAP3K7, TGFB3 and TRAF6 [29-33], confirm-

ing the importance of these two cytokines being promising RA drug targets. The In silico insight into the mechanistic inhibition of TNF- α and IL-1 by resveratrol, is showing the compound to be more efficient in binding to TNF- α . The resveratrol-TNF- α interactions and the number of hydrogen bonds formed are displayed in **Figure 9A**. Resveratrol-TNF- α has a ΔG of -4.95 Kcal/mol. The hydrophobic binding pocket in this case is of following amino acids TRP139, VAL90, GLN38, LEU92, PHE54, LEU24, LEU72, ILE32, ILE74, ARG73, TYR80, VAL81, LEU40, TYR39 and PRO89. **Figure 9B** shows three dimensional mapping of the hydrophobic interaction between resveratrol and TNF- α . The inhibition of this cytokine alone or in tandem with IL-1 is pivotal drug inhibiting strategy for RA. The pathway analysis is conclusively showing the role TNF- α and IL-1 in RA. The in vitro mechanistic approach to validate the binding was beyond the scope of the article; however the immunomodulation was elucidated using oxidative burst assay. The preliminary in vitro results of neutrophil cells with resveratrol showed a dose-dependent effect with >70% inhibition at the highest concentration (10 μ M). Both the In silico and in vitro analysis of resveratrol as a drug for the treatment of rheumatoid arthritis are giving sufficient leads to proceed for further drug development.

Acknowledgements

The protocols/experiments involving the use of human specimens were duly examined and approved by the Hospital Ethics Committee (HEC), Zhujiang hospital of Southern Medical University, Guangzhou (HEC-ZHSMU/2014).

Disclosure of conflict of interest

None.

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References

- [1] Janosy G, Duke O, Poulter L, Panayi G, Bofill M and Goldstein G. Rheumatoid arthritis: a disease of T-lymphocyte/macrophage immunoregulation. *Lancet* 1981; 318: 839-842.
- [2] Klareskog L, Catrina AI and Paget S. Rheumatoid arthritis. *Lancet* 2009; 373: 659-672.
- [3] McInnes IB and O'Dell JR. State-of-the-art: rheumatoid arthritis. *Ann Rheum Dis* 2010; 69: 1898-1906.
- [4] McInnes IB and Schett G. Cytokines in the pathogenesis of rheumatoid arthritis. *Nat Rev Immunol* 2007; 7: 429-442.
- [5] Oliver J and Silman A. Risk factors for the development of rheumatoid arthritis. *Scand J Rheumatol* 2006; 35: 169-174.
- [6] Stark K, Straub RH, Blažičková S, Hengstenberg C and Rovenský J. Genetics in neuroendocrine immunology: implications for rheumatoid arthritis and osteoarthritis. *Ann N Y Acad Sci* 2010; 1193: 10-14.
- [7] Chabaud M, Durand JM, Buchs N, Fossiez F, Page G, Frappart L and Miossec P. Human interleukin-17: A T cell-derived proinflammatory cytokine produced by the rheumatoid synovium. *Arthritis Rheum* 1999; 42: 963-970.
- [8] Nakae S, Nambu A, Sudo K and Iwakura Y. Suppression of immune induction of collagen-induced arthritis in IL-17-deficient mice. *J Immunol* 2003; 171: 6173-6177.
- [9] Hitchon CA, Alex P, Erdile LB, Frank MB, Dozmorov I, Tang Y, Wong K, Centola M and El-Gabalawy HS. A distinct multicytokine profile is associated with anti-cyclical citrullinated peptide antibodies in patients with early untreated inflammatory arthritis. *J Rheumatol* 2004; 31: 2336-2346.
- [10] Eastgate J, Wood N, Di Giovine F, Symons J, Grinlinton F and Duff G. Correlation of plasma interleukin 1 levels with disease activity in rheumatoid arthritis. *Lancet* 1988; 332: 706-709.
- [11] Saxne T, Palladino M, Heinegard D, Talal N and Wollheim F. Detection of tumor necrosis factor α but not tumor necrosis factor β in rheumatoid arthritis synovial fluid and serum. *Arthritis Rheum* 1988; 31: 1041-1045.
- [12] Wood AJ, Olsen NJ and Stein CM. New drugs for rheumatoid arthritis. *N Engl J Med* 2004; 350: 2167-2179.
- [13] den Broeder A, van de Putte L, Rau R, Schattenkirchner M, Van Riel P, Sander O, Binder C, Fenner H, Bankmann Y and Velagapudi R. A single dose, placebo controlled study of the fully human anti-tumor necrosis factor-alpha antibody adalimumab (D2E7) in patients with rheumatoid arthritis. *J Rheumatol* 2002; 29: 2288-2298.
- [14] Zhou W, Cai JF, Yuan F, Ma M and Yin F. In silico targeting of interleukin-6 by natural compounds. *Bangladesh Journal of Pharmacology* 2014; 9: 371-376.
- [15] Khanna D, Sethi G, Ahn KS, Pandey MK, Kunnakkara AB, Sung B, Aggarwal A and Aggar-

In-Silico insight into resveratrol targeting TNF- α and IL-1

- wal BB. Natural products as a gold mine for arthritis treatment. *Curr Opin Pharmacol* 2007; 7: 344-351.
- [16] Bertelli AA and Das DK. Grapes, wines, resveratrol, and heart health. *J Cardiovasc Pharmacol* 2009; 54: 468-476.
- [17] Elliott PJ and Jirousek M. Sirtuins: novel targets for metabolic disease. *Curr Opin Investig Drugs* 2008; 9: 371-378.
- [18] Elmali N, Baysal O, Harma A, Esenkaya I and Mizrak B. Effects of resveratrol in inflammatory arthritis. *Inflammation* 2007; 30: 1-6.
- [19] Molnar V and Garai J. Plant-derived anti-inflammatory compounds affect MIF tautomerase activity. *Int Immunopharmacol* 2005; 5: 849-856.
- [20] Penberthy WT. Pharmacological targeting of IDO-mediated tolerance for treating autoimmune disease. *Curr Drug Metab* 2007; 8: 245-266.
- [21] Chikan NA, Bhavaniprasad V, Anbarasu K, Shabir N and Patel TN. From natural products to drugs for epimutation computer-aided drug design. *Appl Biochem Biotechnol* 2013; 170: 164-175.
- [22] Chikan NA and Vipperla B. KAISO inhibition: an atomic insight. *J Biomol Struct Dyn* 2015; 33: 1794-804.
- [23] He MM, Smith AS, Oslob JD, Flanagan WM, Braisted AC, Whitty A, Cancilla MT, Wang J, Luvogskoy AA, Yoburn JC, Fung AD, Farrington G, Eldredge JK, Day ES, Cruz LA, Cachero TG, Miller SK, Friedman JE, Choong IC and Cunningham BC. Small-molecule inhibition of TNF- α . *Science* 2005; 310: 1022-1025.
- [24] Graves BJ, Hatada MH, Hendrickson WA, Miller JK, Madison VS and Satow Y. Structure of interleukin 1. α . at 2.7- Å . resolution. *Biochemistry* 1990; 29: 2679-2684.
- [25] Inc A. Discovery studio modeling environment. San Diego 2007.
- [26] Morris GM, Huey R, Lindstrom W, Sanner MF, Belew RK, Goodsell DS and Olson AJ. AutoDock4 and AutoDockTools4: Automated docking with selective receptor flexibility. *J Comput Chem* 2009; 30: 2785-2791.
- [27] DeLano WL. The PyMOL molecular graphics system. 2002.
- [28] Hess B, Kutzner C, Van Der Spoel D and Lindahl E. GROMACS 4: algorithms for highly efficient, load-balanced, and scalable molecular simulation. *J Chem Theory Comput* 2008; 4: 435-447.
- [29] Fishman D, Faulds G, Jeffery R, Mohamed-Ali V, Yudkin JS, Humphries S and Woo P. The effect of novel polymorphisms in the interleukin-6 (IL-6) gene on IL-6 transcription and plasma IL-6 levels, and an association with systemic-onset juvenile chronic arthritis. *J Clin Invest* 1998; 102: 1369.
- [30] Hulin-Curtis S, Sharif M, Bidwell J and Perry M. Evaluation of NFKB1A variants in patients with knee osteoarthritis. *Int J Immunogenet* 2013; 40: 272-279.
- [31] Rabelo Fde S, da Mota LM, Lima RA, Lima FA, Barra GB, de Carvalho JF and Amato AA. The Wnt signaling pathway and rheumatoid arthritis. *Autoimmun Rev* 2010; 9: 207-210.
- [32] Litton M, Lindroos E and Klareskog L. Cytokine production in synovial tissue of mice with collagen-induced arthritis (CIA). *Clin Exp Immunol* 1997; 107: 485-493.
- [33] Pauley KM, Satoh M, Chan AL, Bubb MR, Reeves WH and Chan EK. Upregulated miR-146a expression in peripheral blood mononuclear cells from rheumatoid arthritis patients. *Arthritis Res Ther* 2008; 10: R101.