

## Review Article

# A brief review of perianal paget disease

Mingbo Tang<sup>1</sup>, Xinliang Gao<sup>1</sup>, Lizhe Wang<sup>2</sup>, Wei Liu<sup>1</sup>, Jiannan Li<sup>3</sup>

Departments of <sup>1</sup>Thoracic Surgery, <sup>2</sup>Pediatric Oncology, The First Hospital of Jilin University, Changchun, Jilin, P. R. China; <sup>3</sup>Department of General Surgery, The Second Hospital of Jilin University, Changchun, Jilin, P. R. China

Received May 24, 2020; Accepted August 11, 2020; Epub October 15, 2020; Published October 30, 2020

**Abstract:** Paget's disease (PD) is a kind of malignant tumor that is usually localized within the epidermis. PD can be divided into mammary PD (MPD) and extramammary PD (EMPD). EMPD is rare, but the actual incidence is not known. Perianal PD (PPD) refers to EMPD that is located within 6 cm of the anal orifice and below the pectinate line. In this study, we summarize the pathogenesis, clinical characteristics, diagnosis, and treatment of PPD. Pubmed and Web of Science data-bases were used to search for the relevant studies, and the key words were "EMPD" and "PPD". The symptoms of PPD are similar to those of other benign skin diseases, and so the diagnosis of PPD is always delayed. Histopathology is necessary for the diagnosis of PPD. Many treatment methods have been used for PPD, but surgical resection remains the treatment of choice. The rarity of PPD has hampered further research.

**Keywords:** PD, EMPD, PPD, histopathology, surgery

## Introduction

Paget's disease (PD) is a rare type of cancer that arises in the epidermis. It is classified as mammary PD (MPD) and extramammary PD (EMPD). MPD arises in the epidermis of the nipple or the areola of the breast [1], and was first reported by James Paget in 1874 [2]. In 1889, Radcliffe Crocker described EMPD in a male patient in whom the disease presented with eczema-like lesions on the skin of the penis and scrotum [3]. EMPD can occur at any site where apocrine glands are present, e.g., the vulva, perineum, and perianal region [4]. Perianal PD (PPD), which was first described by Darier and Couillaud [5, 6], refers to lesions which are located below the pectinate line and within 6 cm of the anal orifice. Patients with PPD present with well-defined erythematous lesions and invariably complain of itching [7]. Because the symptoms of PPD are similar to those of many benign skin diseases, the diagnosis of PPD is always delayed [8].

The precise incidence and prevalence of EMPD are unknown. PD accounts for about 4% of all breast cancers, while EMPD accounts for about 6.5% of all skin diseases [9, 10]. The most common site for EMPD is the vulva, followed by the perianal area, penis, scrotum, and groin area [11]. The incidence of PPD is hard to estimate,

but it is believed that PPD accounts for about 20% of EMPD and 6% of PD [12]. PPD is often accompanied by other cancers, such as ovarian and colorectal cancers [13].

In this study, we summarize the pathogenesis, clinical characteristics, diagnosis, and treatment of PPD. This information will be helpful for early diagnosis and proper treatment of PPD.

## Pathogenesis of PPD

EMPD may originate from three different cell types [8]: 1) Derived from the apocrine adenocarcinoma or exocrine adenocarcinoma, especially the sweat gland; 2) Originated from the adenocarcinoma in the epidermis, which is also called intraepithelial neoplasia; 3) Derived from adenocarcinoma of other organs spread to the epidermis.

PPD can be divided into two types: primary cutaneous PPD and secondary dermal-derived PPD [14]. Primary cutaneous PPD is a type of epithelial adenocarcinoma and always presents with intraepithelial infiltration. Its precursor cells appear to be the undifferentiated pluripotent cells of the epidermis or skin appendages. Epidermal cells are also considered to be precursors of Paget cells. Primary cutaneous PPD is rarely invasive, and so metastasis is

## Perianal paget disease



**Figure 1.** Different presentations of perianal Paget disease: A. Erythematous skin lesion in the perianal area with surrounding lichenification [15]. B. A thickened plaque like area in the perineal region [16].

uncommon [14]. Secondary dermal-derived PPD originates from the adenocarcinoma tissues of other organs. For example, rectal adenocarcinoma and urinary tract cancer cells may spread to the perianal epidermis and lead to secondary dermal-derived PPD. When secondary dermal-derived PPD is diagnosed, further examination must be conducted to identify the primary malignancy [14].

### Clinical characteristics

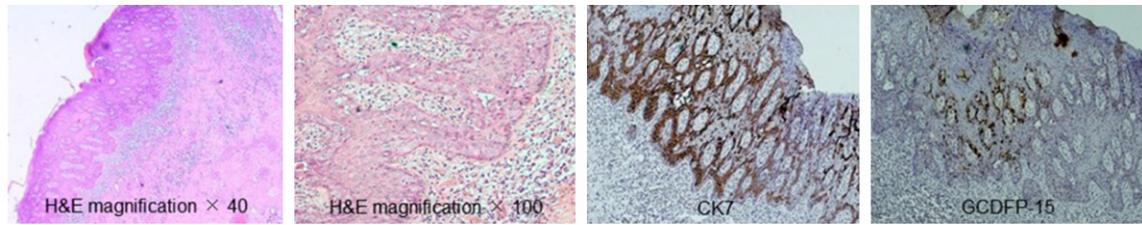
Because EMPD is a slow-growing intraepithelial tumor, its clinical manifestations are often nonspecific. PPD patients invariably have rash, accompanied by itching and pain in the affected area. In addition, some patients complain about hematochezia, perianal swelling, and change in bowel habits. The skin color in the affected area can vary from pink to dark red, and large lesions may present with multiple colors. There may be scales, exudates, patchy erosions, or white spots on the surface of the lesions. Severe lesions may be irregular, with unclear boundaries. Due to centrifugal growth, large lesions may completely involve the anal and genital area, leading to the formation of polygonal boundaries [9]. Zeng *et al.* reported a rare case of PPD that presented as an erythematous skin lesion in the perianal area with surrounding lichenification (**Figure 1A**) [15]. The patient complained about pruritus, but had no pain or bleeding. Another patient presented with a butterfly-shaped thickened plaque in the perineal region, with sparing of the anal opening (**Figure 1B**) [16]. This patient had pruritus and mild pain in the perianal area and also

complained about a “leathery” feel of the affected skin.

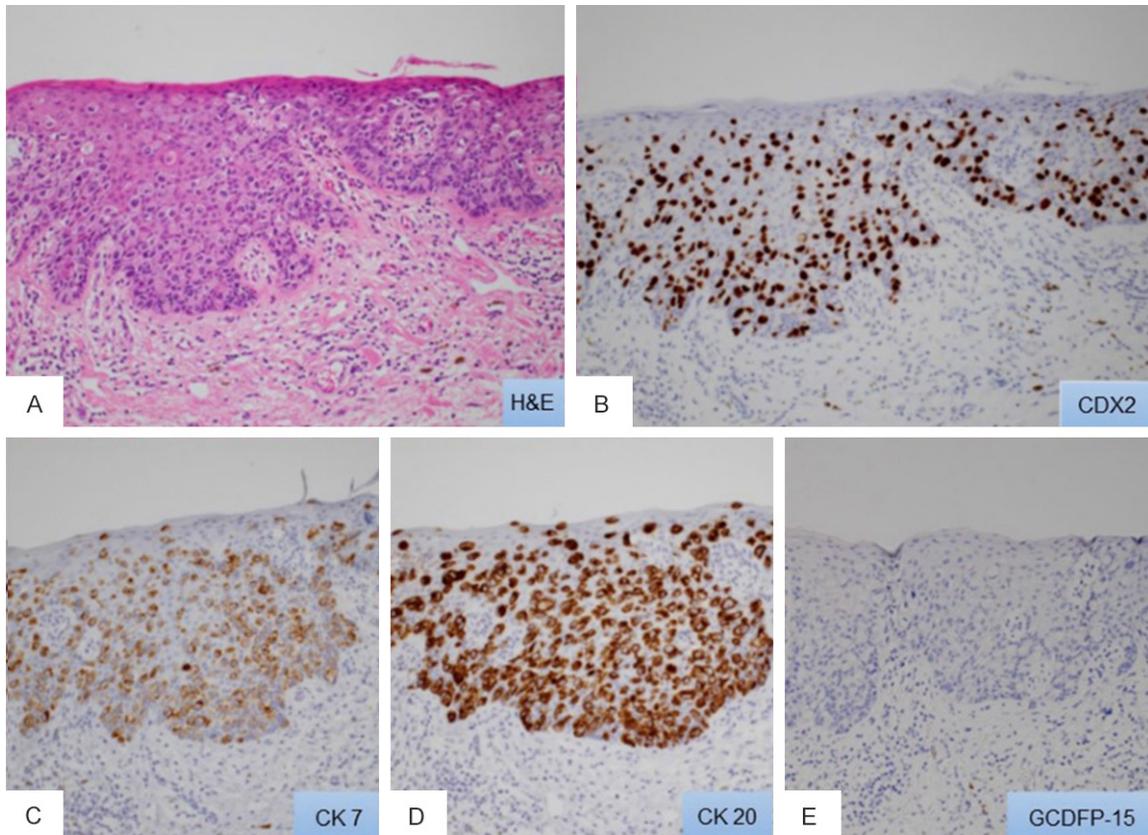
### Diagnosis of PPD

The diagnosis of PPD is based on histopathological and immunohistochemical analysis [9]. There are differences between primary cutaneous PPD and secondary dermal-derived PPD in the histopathological and immunohistochemical features [17]. It is important to distinguish between the two types of PPD because the surgical methods and prognosis are different. Secondary dermal-derived PPD mostly originates from anal cancer or low rectal cancer and carries a worse prognosis. Under the microscope, primary cutaneous PPD typically shows uniformly distributed tumor cells, with only occasional glandular cavities. Whereas secondary dermal-derived PPD is characterized by irregularly arranged tumor cells and a relatively greater number of glandular cavities [18]. Immunohistochemistry of cytokeratin (CK)-7, CK-20, and gross cystic disease fluid protein (GCDFP-15) can help in differentiating different types of EMPDs. EMPD tissues originating from urothelial carcinoma will be positive for CK7 and CK20 and negative for GCDFP-15. EMPD derived from the urinary tract is commonly positive for GCDFP-15 [19]. PPD derived from rectal and anal cancer is usually positive for CK-20 and negative for CK7 [20], although some cases may be positive for CK7 and negative for CK20 [21]. Pandey *et al.* reported a case of histopathology-confirmed primary cutaneous PPD that was positive for CK7 and GCDFP-15 (**Figure 2**) [16].

## Perianal paget disease



**Figure 2.** Histological features of primary cutaneous perianal Paget disease [16].



**Figure 3.** Histological findings of perianal Paget disease [26]. (A) Paget cells with clear cytoplasm and large pleomorphic nuclei (hematoxylin-eosin; magnification  $\times 20$ ). (B-D) Positive immunohistochemical staining for CDX2 (B), CK7 (C), and CK20 (D), and negative staining for GCDFFP-15 (E) (magnification  $\times 20$ ).

Carcinoembryonic antigen (CEA) is also a useful marker of urothelium-derived PPD. However, some cases of rectal and anal cancer-derived PPD may also be negative for CEA. Therefore, as with CK7 and CK20, CEA staining alone is insufficient for definite diagnosis of urothelium-derived EMPD.

Caudal-related homeobox gene nuclear transcription factor-2 (CDX2) is a gene involved in the regulation of the proliferation/differentiation of intestinal cells. Positive staining for

CDX2 is common in both primary and metastatic rectal cancer [22-24]. Nisi *et al.* found primary EMPD to be positive for CK7 and negative for CK20 and CDX2, and secondary EMPD to be positive for CDX2 [25]. CDX2 is therefore considered a sensitive marker for rectal- and anal-derived EMPD. In one report of anal gland carcinoma *in situ* with pagetoid spread [26], histological examination showed Paget cells with clear cytoplasm, and immunohistochemistry was positive for CK7, CK20, and CDX2 and negative for GCDFFP-15 (**Figure 3**).

Recently, mucin staining has been proposed as a sensitive method for diagnosis of EMPD. Mucin is a high-molecular-weight glycoprotein with nine subtypes (MUC1-9) which are differentially expressed in different types of tissues [27]. MUC1 is often found on the apical surface of the glandular epithelium and has recently been shown to be a very sensitive marker of EMPD [28, 29]. Some researchers have reported the typical patterns of mucin staining in EMPD. Kuan *et al.* reported three cases of rectal adenocarcinoma-derived PPD that stained positive for MUC2 [30]. Kondo *et al.* found diffusely positive MUC2 staining in two of three patients with primary PPD [27]. Liegl *et al.* reported 23 patients with EMPD who had positive MUC1 staining [29]. Yoshii *et al.* reported 36 patients with EMPD, 2 of whom had PPD [28]. Both PPD patients showed negative staining for MUC2 and no underlying adenocarcinoma was found in the PPD tissues. Some researchers have reported MUC5AC in some EMPD tissues [28, 30].

It is necessary to differentiate PPD from Bowen disease, contact dermatitis, mossy lesions, psoriasis, melanoma, perianal Crohn disease, mycosis fungoides, squamous cell carcinoma, and femoral hernia. In Bowen disease, the lesion presents as irregular erythema with a clear boundary and varying degrees of scaling, desquamation, and even erosion and exudation. Histopathology shows atypical hyperplasia with pleomorphic nuclei. Some lesions can progress to invasive cancer. Bowen disease may be related to chronic sun damage, immune abnormalities, chronic irritation, human papilloma virus infection, and trauma [31, 32]. Matsumoto *et al.* reported that whereas Paget cells were positive for CEA, CK7, and CK8, the atypical keratinocytes of Bowen disease were negative for CEA but positive for CK7 and CK8 [33]. Thus, CEA could be used to distinguish EMPD from Bowen disease. Pigment present in the cytoplasm of some EMPD tumor cells could lead to suspicion of malignant melanoma. However, the tumor cells of Paget-like malignant melanoma are atypical or clustered and are rich in melanin. On immunohistochemistry, malignant melanoma cells are positive for S-100, HMB-45, melan-A, and MART-1, whereas Paget cells are negative for all of them. Differentiation of EMPD from other diseases may require direct mycological examination

and mycological culture in addition to histopathological analysis.

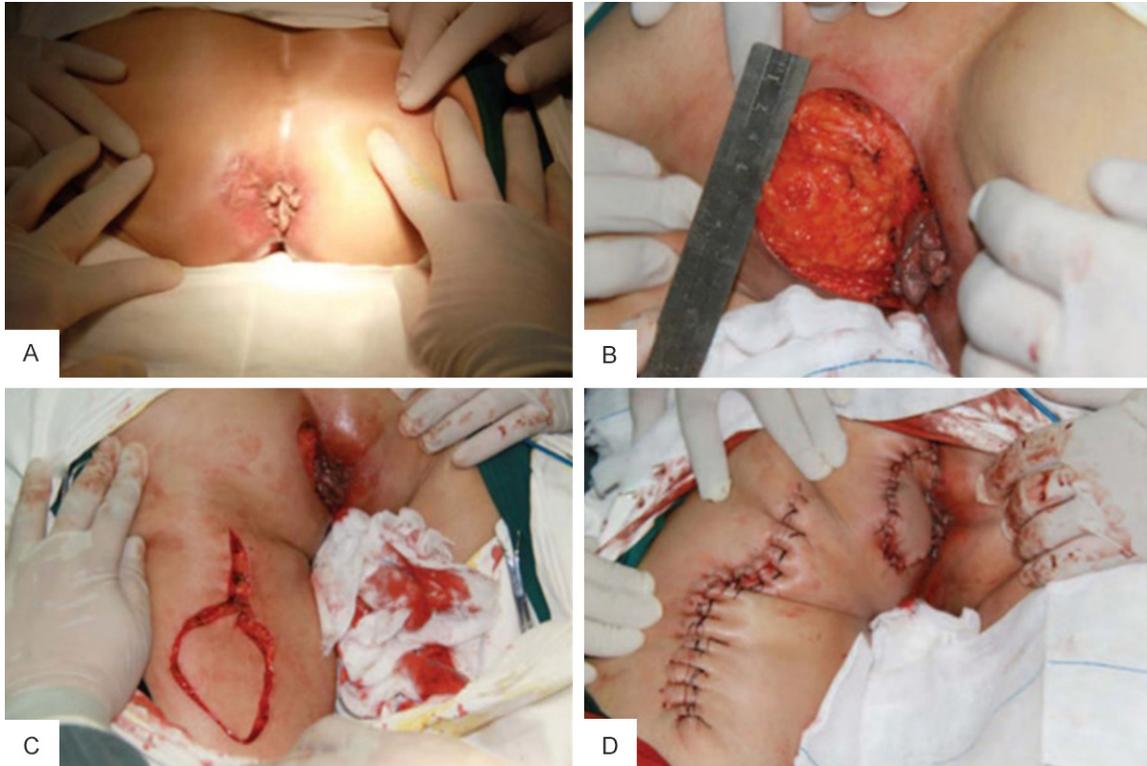
### Treatment of PPD

Treatment methods for PPD include surgery, photodynamic treatment, 5% imiquimod cream application, CO<sub>2</sub> laser ablation, radiotherapy, and chemotherapy.

Surgical treatment is applicable for epidermal/intradermal PPD in the perineum, scrotum, or vulva [3, 34, 35]. Most researchers believe that wide local excision of the visible lesion is adequate for PPD treatment. However, excision results in large tissue defects. In most cases, primary suture or skin grafting will not be appropriate, and flap reconstruction surgery will be required. Importantly, the reconstruction of the perianal defect has to ensure the preservation of anal function and satisfactory cosmetic results. Good results have been reported with bilateral musculocutaneous flaps from the gluteal or thigh regions and with “V-Y” island flaps [36, 37]. Kishi *et al.* reconstructed anal and perianal areas using a posterior thigh trilobed flap and reported good results [38]. Shen *et al.* reported a female PPD patient who underwent wide local excision of the lesion followed by posterior thigh reconstruction [39]. There were no complications and the patient achieved satisfactory bowel control (**Figure 4**). The posterior thigh flap has certain advantages for reconstruction of perianal defects. The donor site is close to the defect and flap transfer is easy. In addition, the flap has good blood supply, which significantly increases the survival rate [40]. In severe PPD, colostomy may be required before extensive local excision. Some researchers recommend that colostomy should be performed when more than half of the perianal area needs to be removed, or when the resection radius is >3 cm [36].

Literature review shows that the recurrence rate of primary EMPD after standard extensive local resection is as high as 60% [41]. Recurrence may be local, in the lymph nodes, or at a distant location [42]. The determination of the resection margin during surgery is a major challenge. Intraoperative frozen section analysis can help to a certain extent. However, because of the multicentric nature of PPD and the limited time for intraoperative analysis, the false negative rate of frozen section analysis is

## Perianal paget disease



**Figure 4.** Surgical treatment of perianal Paget disease [39]. A. The lesion in the left perianal area measures 3 × 3 cm and shows surrounding lichenification. B. The defect is 7 × 6 cm in size. C. Posterior thigh flap is designed according to the defect in the perianal region. D. The defect after flap reconstruction.

about 40% [43]. To address this problem, Mecker developed a method for resection of PPD in stages [44]. In stage 1 (contoured margin excision), the margin of the lesion is first marked. Then an outline is drawn 1-2 cm away from the marked margin and the direction is determined. Under local or general anesthesia, 2-3 mm of tissue at the incision site is removed and sent for pathological analysis. The wound is sutured and the patient is discharged from hospital. If the pathology report indicates a positive margin, the patient is readmitted and the affected edge is resected. The first stage can be repeated as needed. Stage 2 (central resection and reconstruction) is undertaken once the margin is confirmed to be negative. The wound is reconstructed by plastic surgery. The Mecker method not only guarantees complete tumor resection, but also provides a clear boundary. It is a convenient method to avoid excessive resection and consequent loss of function.

Although EMPD progresses slowly, local and distant metastasis can occur through direct

dermal invasion and lymphatic spread. Treatment delay may cause serious harm. Standard surgical resection may not be sufficient because PPD is invasive and prone to recur. Because visual examination cannot identify the extent of invasion, it is difficult to determine the optimal surgical resection range. Mohs micro-surgery (MMS) may be useful. MMS is a tissue-retaining surgery used for treatment of recurrent, invasive, high-risk skin cancers. In MMS, the surgeon can directly observe the edge of the lesion through a microscope, which makes optimal resection more likely and thus reduces the risk of local recurrence. A retrospective study has suggested that MMS may be better than standard local extensive resection, with a recurrence rate as low as 16% [41].

Photodynamic therapy (PDT) has also been applied for the treatment of EMPD. PDT is based on a photochemical reaction caused by the combination of a pro-neutral photosensitizer and light [45, 46]. A topical photosensitizer is selectively localized to the tumor and then activated by light, resulting in destruction of the

tumor tissue without damaging healthy tissue. Aminolevulinic acid (ALA) or its methyl ester (M-ALA) are the most commonly used photosensitizers. Recent studies have shown good response rates and symptom control with PDT [47-49]. A phase II prospective clinical trial found that although M-ALA-based PDT does not cure PPD, it does help improve life quality and is better than surgical resection [50].

Imiquimod (5%) cream has been used alone and in combination with other methods for treatment of PPD. Imiquimod 5% cream is an immune response modifier and stimulator that is approved for the treatment of genital and perianal fistulas [51]. Zempognoa *et al.* reported two cases of PPD who were successfully treated with imiquimod 5% cream [52]. In another study, topical imiquimod treatment of PPD in a patient without primary gastrointestinal neoplasia resulted in complete regression of the lesion, and no recurrence was noted over a 12-month follow-up period [53]. The most common side effects of imiquimod 5% cream are local skin irritation, erythema, and erosion, which may occur at 4-6 weeks after the initiation of imiquimod treatment [51]. These side effects are dose-dependent and can be alleviated by decreasing the frequency of topical application or with the withdrawal time prolonged [51].

Carbon dioxide (CO<sub>2</sub>) laser ablation has been used to treat PPD in the vulva as it can preserve vulvar anatomy. However, the treatment is painful and local recurrence rates are very high [54]. In one study, the recurrence rates of PPD after local surgery, CO<sub>2</sub> laser ablation, and extensive local excision were 56%, 33%, and 23%, respectively [55].

Radiation therapy is another method for treatment of EMPD. It has been used as the definitive treatment for PPD and also for prevention of postoperative local recurrence. There are several reports on the effectiveness of radiation therapy in noninvasive PPD [56-58]. Yanagi *et al.* used skin biopsy before and after radiotherapy to confirm the effectiveness of radiation therapy [59]. The recommended radiation dose is 40-50 Gy and lower doses are associated with higher risk of disease recurrence [60]. The main side effects after radiation therapy are local telangiectasia and fibrosis.

Systemic chemotherapy for EMPD has been minimally advanced. Various chemotherapy regimens have been reported for the treatment of EMPD, including low-dose 5-fluorouracil (5-FU) and cisplatin [61], vinorelbine and cisplatin [62], 5-FU and leucovorin [63], and a combination of mitomycin C, etoposide, and cisplatin [64]. Hallak *et al.* reported good results in patients with PPD treated with oxaliplatin-assisted chemotherapy after extensive local excision and abdominoperineal resection [65]. The combination of 5-FU local chemotherapy and local surgery has also been used for treatment of EMPD [66]. Systemic chemotherapy can be used as monotherapy or in combination with other methods for EMPD patients with distant organ metastases.

### Prognosis of PPD

The depth of tumor invasion and the number of metastatic lymph nodes are closely related to the prognosis of PPD [67]. Metastasis is rare in EMPD, but when it does occur, the most common sites are lymph nodes and bones. PPD may be more aggressive than genital and perineal EMPD. One study showed significantly lower disease-specific survival in PPD associated with rectal/anal cancer than in PPD not associated with rectal/anal cancer [68]. However, this finding was not confirmed in multivariate analysis [68].

### Conclusions

Because of the rarity of PPD, all studies so far have had small sample sizes. Most of the researchers recommend surgical resection as the primary treatment method. The effectiveness of chemotherapy and radiation therapy is uncertain. PPD is more common among Asians, and Chinese clinicians need to be alerted to the possibility of PPD when a patient presents with abnormal perianal skin. Perianal skin biopsy should be considered for perianal rashes not responding to the usual treatments. Moreover, as PPD can be asymptomatic, all anorectal biopsies should be examined histologically. Because of the risk of invasive anal/perineal cancer in patients with PPD, close follow-up is required after resection, regardless of the margin status. Physicians should be aware of the risks of PPD-related cancer and should not exclude these risks during treatment.

**Acknowledgements**

This study was financially supported by the Financial Department of Jilin Province (Grant No. 2019SCZT045).

**Disclosure of conflict of interest**

None.

**Address correspondence to:** Wei Liu, Department of Thoracic Surgery, The First Hospital of Jilin University, Changchun, Jilin, P. R. China. Tel: +86-13596083366; E-mail: liuweihospital@126.com; Jiannan Li, Department of General Surgery, The Second Hospital of Jilin University, Changchun, Jilin, P. R. China. Tel: +86-13894897951; E-mail: jnli@ciac.ac.cn

**References**

[1] Morris CR and Hurst EA. Extramammary paget disease: a review of the literature-part i: history, epidemiology, pathogenesis, presentation, histopathology, and diagnostic work-up. *Dermatol Surg* 2020; 46: 151-158.

[2] Lloyd J and Flanagan AM. Mammary and extramammary Paget's disease. *J Clin Pathol* 2000; 53: 742-749.

[3] Simonds RM, Segal RJ and Sharma A. Extramammary Paget's disease: a review of the literature. *Int J Dermatol* 2019; 58: 871-879.

[4] Mengjun B, Zheng-Qiang W and Tasleem MM. Extramammary Paget's disease of the perianal region: a review of the literature emphasizing management. *Dermatol Surg* 2013; 39: 69-75.

[5] Marchesa P, Fazio VW, Oliart S, Goldblum JR, Lavery IC and Milsom JW. Long-term outcome of patients with perianal Paget's disease. *Ann Surg Oncol* 1997; 4: 475-480.

[6] Tulchinsky H, Zmora O, Brazowski E, Goldman G and Rabau M. Extramammary Paget's disease of the perianal region. *Colorectal Dis* 2004; 6: 206-209.

[7] dos Santos JS, Bonafe GA, Pereira JA, Kanno DT, Real Martinez CA and Ortega MM. Rare perianal extramammary Paget disease successfully treated using topical Imiquimod therapy. *BMC Cancer* 2018; 18: 921.

[8] Gaertner WB, Hagerman GF, Goldberg SM and Finne CO 3rd. Perianal Paget's disease treated with wide excision and gluteal skin flap reconstruction: report of a case and review of the literature. *Dis Colon Rectum* 2008; 51: 1842-1845.

[9] Lopes Filho LL, Lopes IM, Lopes LR, Enokihara MM, Michalany AO and Matsunaga N. Mam-

mary and extramammary Paget's disease. *An Bras Dermatol* 2015; 90: 225-231.

[10] Kanitakis J. Mammary and extramammary Paget's disease. *J Eur Acad Dermatol Venereol* 2007; 21: 581-590.

[11] Chanda JJ. Extramammary Paget's disease: prognosis and relationship to internal malignancy. *J AM Acad Dermatol* 1985; 13: 1009-1014.

[12] Rao SD and Govindarajan M. Extramammary Paget's disease of the perianal region. *Indian J Surg* 2017; 79: 360-362.

[13] Kim CW, Kim YH, Cho MS, Min BS, Baik SH and Kim NK. Perianal Paget's disease. *Ann Coloproctol* 2014; 30: 241-244.

[14] Chaudhuri SP and Smoller BR. Extramammary Paget's disease: diagnosis and disease pattern. *Cutis* 1992; 50: 195-196.

[15] Zeng D, Chen J, Zhu B, Li J, Wu H and Ma D. Laparoscopic extralevator abdominoperineal excision for the treatment of perianal Paget's disease: a case report. *Medicine* 2019; 98: e15243.

[16] Pandey A, Singh P, Mishra R and Gupta A. Primary perianal extramammary Paget's disease: case report with review. *J Dermatol Dermatol Surg* 2016; 20: 152-155.

[17] Ackerman AB. Differential diagnosis in dermatology. *JAMA* 1983; 249: 882-882.

[18] Wong AY, Rahilly MA, Adams W and Lee CS. Mucinous anal gland carcinoma with perianal Pagetoid spread. *Pathol* 1998; 30: 1-3.

[19] Brown HM and Wilkinson EJ. Uroplakin-III to distinguish primary vulvar Paget disease from Paget disease secondary to urothelial carcinoma. *Hum Pathol* 2002; 33: 545-548.

[20] Ohnishi T and Watanabe S. The use of cytokeratins 7 and 20 in the diagnosis of primary and secondary extramammary Paget's disease. *Brit J Dermatol* 2000; 142: 243-247.

[21] Zhang PJ, Shah M, Spiegel GW and Brooks JJ. Cytokeratin 7 immunoreactivity in rectal adenocarcinomas. *Appl Immunohisto M M* 2003; 11: 306-310.

[22] Lora V and Kanitakis J. CDX2 expression in cutaneous metastatic carcinomas and extramammary Paget's disease. *Anticancer Res* 2009; 29: 5033-5037.

[23] Barbareschi M, Murer B, Colby TV, Chilosi M, Macri E, Loda M and Doglioni C. CDX-2 homeobox gene expression is a reliable marker of colorectal adenocarcinoma metastases to the lungs. *AM J Surg Pathol* 2003; 27: 141-149.

[24] Yatabe Y, Koga T, Mitsudomi T and Takahashi T. CK20 expression, CDX2 expression, K-ras mutation, and goblet cell morphology in a subset of lung adenocarcinomas. *J Pathol* 2004; 203: 645-652.

[25] De Nisi MC, D'Amuri A, Toscano M, Lalinga AV, Pirtoli L and Miracco C. Usefulness of CDX2 in

## Perianal paget disease

- the diagnosis of extramammary Paget disease associated with malignancies of intestinal type. *Br J Dermatol* 2005; 153: 677-679.
- [26] Ishioka K, Koyama F, Inoue T, Obara S, Nakamoto T, Sasaki Y, Nakamura Y, Takeda M, Ohbayashi C, Kuwahara M and Sho M. Anal gland adenocarcinoma in situ with pagetoid spread: a case report. *Surg Case Rep* 2018; 4: 63.
- [27] Kondo Y, Kashima K, Daa T, Fujiwara S, Nakayama I and Yokoyama S. The ectopic expression of gastric mucin in extramammary and mammary Paget's disease. *AM J Surg Pathol* 2002; 26: 617-623.
- [28] Yoshii N, Kitajima S, Yonezawa S, Matsukita S, Setoyama M and Kanzaki T. Expression of mucin core proteins in extramammary Paget's disease. *Pathol Int* 2002; 52: 390-399.
- [29] Liegl B, Leibl S, Gogg-Kamerer M, Tessaro B, Horn LC and Moinfar F. Mammary and extramammary Paget's disease: an immunohistochemical study of 83 cases. *Histopathol* 2007; 50: 439-447.
- [30] Kuan SF, Montag AG, Hart J, Krausz T and Recant W. Differential expression of mucin genes in mammary and extramammary Paget's disease. *AM J Surg Pathol* 2001; 25: 1469-1477.
- [31] Caca-Biljanovska N, Arsovska-Bezhoska I and V'Lckova-Laskoska M. Giant Bowen's disease on the face: case report and review of the literature. *Open Access Maced J Med Sci* 2019; 7: 606-609.
- [32] Narahira A, Yanagi T, Kitamura S, Hata H and Shimizu H. Dermoscopic features of genital pigmented Bowen's disease: report of a case and review of the published work. *J Dermatol* 2019; 46: E390-E391.
- [33] Matsumoto M, Ishiguro M, Ikeno F, Ikeda M, Kamijima R, Hirata Y, Saruta T and Kodama H. Combined bowen disease and extramammary paget disease. *J Cutan Pathol* 2007; 34: 47-51.
- [34] Shutze WP and Gleysteen JJ. Perianal Paget's disease. Classification and review of management: report of two cases. *Dis Colon Rectum* 1990; 33: 502-507.
- [35] Chung PH, Leong JY and Voelzke BB. Surgical experience with genital and perineal extramammary Paget's disease. *Urol* 2019; 128: 90-95.
- [36] Hassan I, Horgan AF and Nivatvongs S. V-Y Island flaps for repair of large perianal defects. *AM J Surg* 2001; 181: 363-365.
- [37] Mardini S, Tsai FC and Wei FC. The thigh as a model for free style free flaps. *Clin Plast Surg* 2003; 30: 473-480.
- [38] Kishi K, Nakajima H, Imanishi N and Nakajima T. Anal and perianal reconstruction after extramammary Paget's disease using a posterior thigh flap with a thin square wing. *J Plast Reconstr Aes* 2010; 63: 1353-1356.
- [39] Shen K, Luo H, Hu J and Xie Z. Perianal Paget disease treated with wide excision and thigh skin flap reconstruction: a case report and review of literature. *Medicine* 2018; 97: e11638.
- [40] Stavrou M, Martin L, El-Madani F, Naik V, Papanastasiou S and Gupta S. Perianal Paget's disease-report of a rare case. *Int J Surg Case Rep* 2012; 3: 483-485.
- [41] Hendi A, Brodland DG and Zitelli JA. Extramammary Paget's disease: surgical treatment with Mohs micrographic surgery. *J AM Acad Dermatol* 2004; 51: 767-773.
- [42] Minicozzi A, Borzellino G, Momo R, Steccanella F, Pitoni F and de Manzoni G. Perianal Paget's disease: presentation of six cases and literature review. *Int J Colorectal Dis* 2010; 25: 1-7.
- [43] Gunn RA and Gallager HS. Vulvar Paget's disease: a topographic study. *Cancer* 1980; 46: 590-594.
- [44] Moller MG, Lugo-Baruqui JA, Milikowski C and Salgado CJ. Staged marginal contoured and central excision technique in the surgical management of perianal Paget's disease. *AM J Surg* 2014; 207: 485-492.
- [45] Soler AM, Warloe T, Berner A and Giercksky KE. A follow-up study of recurrence and cosmesis in completely responding superficial and nodular basal cell carcinomas treated with methyl 5-aminolaevulinate-based photodynamic therapy alone and with prior curettage. *Brit J Dermatol* 2001; 145: 467-471.
- [46] Vinciullo C, Elliott T, Francis D, Gebauer K, Spelman L, Nguyen R, Weightman W, Sheridan A, Reid C, Czarnecki D and Murrell D. Photodynamic therapy with topical methyl aminolaevulinate for 'difficult-to-treat' basal cell carcinoma. *Brit J Dermatol* 2005; 152: 765-772.
- [47] Nardelli AA, Stafinski T and Menon D. Effectiveness of photodynamic therapy for mammary and extra-mammary Paget's disease: a state of the science review. *BMC Dermatol* 2011; 11: 13-13.
- [48] Li Q, Gao T, Jiao B, Qi X, Long HA, Qiao H, Wang L, Lv Y, Hu X, Liao W, Wang S and Li C. Long-term follow-up of in situ extramammary Paget's disease in asian skin types IV/V treated with photodynamic therapy. *Acta Derm-Venereol* 2010; 90: 159-164.
- [49] Housel JP, Izikson L and Zeitouni NC. Noninvasive extramammary Paget's disease treated with photodynamic therapy: case series from the roswell park cancer institute. *Dermatol Sur* 2010; 36: 1718-1724.
- [50] Fontanelli R, Papadia A, Martinelli F, Lorusso D, Grijuela B, Merola M, Solima E, Ditto A and

## Perianal paget disease

- Raspagliesi F. Photodynamic therapy with M-ALA as non surgical treatment option in patients with primary extramammary Paget's disease. *Gyn Oncol* 2013; 130: 90-94.
- [51] Navi D and Huntley A. Imiquimod 5 percent cream and the treatment of cutaneous malignancy. *Dermatol Online J* 2004; 10: 4.
- [52] Zampogna JC, Flowers FP, Roth WI and Hassenein AM. Treatment of primary limited cutaneous extramammary Paget's disease with topical imiquimod monotherapy: two case reports. *J AM Acad Dermatol* 2002; 47: S229-S235.
- [53] Vereecken P, Awada A, Ghanem G, da Costa CM, Larsimont D, Simoens C, da Costa PM and Hendlisz A. A therapeutic approach to perianal extramammary Paget's disease: topical imiquimod can be useful to prevent or defer surgery. *Med Sci Monitor* 2007; 13: CS75-CS77.
- [54] Valentine BH, Arena B and Green E. Laser ablation of recurrent Paget's disease of vulva and perineum. *J Gyn Surg* 1992; 8: 21-24.
- [55] Louis-Sylvestre C, Haddad B and Paniel BJ. Paget's disease of the vulva: results of different conservative treatments. *Eur J Obst Gyn R B* 2001; 99: 253-255.
- [56] Dilme-Carreras E, Iglesias-Sancho M, Marquez-Balbas G, Sola-Ortigosa J and Umbert-Millet P. Radiotherapy for extramammary Paget disease of the anogenital region. *J AM Acad Dermatol* 2011; 65: 192-194.
- [57] Adkisson CD and Landmann RG. A hemorrhoid by any other name. *Mayo Clin Proc* 2011; 86: e25-e25.
- [58] Secco GB, Lapertosa G, Sertoli MR, Scarpati D and Riboli EB. Perianal Paget's disease: case report of an elderly patient treated with polychemotherapy and radiotherapy. *Tumori* 1984; 70: 381-383.
- [59] Yanagi T, Kato N, Yamane N and Osawa R. Radiotherapy for extramammary Paget's disease: histopathological findings after radiotherapy. *Clin Exp Dermatol* 2007; 32: 506-508.
- [60] Moreno-Arias GA, Conill C, Castells-Mas A, Arenas M and Grimalt R. Radiotherapy for genital extramammary Paget's disease in situ. *Dermatol Surg* 2001; 27: 587-590.
- [61] Matsushita S, Yonekura K, Mera K, Kawai K and Kanekura T. Successful treatment of metastatic extramammary Paget's disease with S-1 and docetaxel combination chemotherapy. *J Dermatol* 2011; 38: 996-998.
- [62] Takahagi S, Noda H, Kamegashira A, Madokoro N, Hori I, Shindo H, Mihara S and Hide M. Metastatic extramammary Paget's disease treated with paclitaxel and trastuzumab combination chemotherapy. *J Dermatol* 2009; 36: 457-461.
- [63] Mochitomi Y, Sakamoto R, Gushi A, Hashiguchi T, Mera K, Matsushita S, Nishi M, Kanzaki T and Kanekura T. Extramammary Paget's Disease/carcinoma successfully treated with a combination chemotherapy: report of two cases. *J Dermatol* 2005; 32: 632-637.
- [64] Watanabe Y, Hoshiai H, Ueda H, Nakai H, Obata K and Noda K. Low-dose mitomycin C, etoposide, and cisplatin for invasive vulvar Paget's disease. *Int J Gynecol Cancer* 2002; 12: 304-307.
- [65] Al Hallak MN and Zouain N. Extramammary perianal Paget's disease. *Case Rep Gastroen* 2009; 3: 332-337.
- [66] Zollo JD and Zeitouni NC. The roswell park cancer institute experience with extramammary Paget's disease. *Brit J Dermatol* 2000; 142: 59-65.
- [67] Ito T, Kaku Y, Nagae K, Nakano-Nakamura M, Nakahara T, Oda Y, Hagihara A, Furue M and Uchi H. Tumor thickness as a prognostic factor in extramammary Paget's disease. *J Dermatol* 2015; 42: 269-275.
- [68] Karam A and Dorigo O. Treatment outcomes in a large cohort of patients with invasive extramammary Paget's disease. *Gynecol Oncol* 2012; 125: 346-351.