Original Article

Chronic wound treatment with high-density nanofat grafting combined with negative pressure wound therapy

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Abstract: Negative pressure wound therapy is an essential technological advancement in wound care and stem cell therapy is the most promising treatment for chronic wounds. Nanofat grafting, an adipose-derived stem cell-based cytotherapy, has shown therapeutic potential for various conditions. However, the effects of negative pressure wound therapy combined with nanofat grafting on wound healing have not been investigated. In this study, high-density nanofat grafting combined with negative pressure wound therapy was used to treat patients with chronic wounds, while negative pressure wound therapy alone was used as the control. Data were analyzed with independent sample t-tests. A comparative study of wound healing rates, post-surgery, and histological changes revealed that average wound healing rates in the high-density nanofat combined with negative pressure wound therapy group were 26.50% ± 9.51%, compared to 12.02% ± 4.20% in the negative pressure wound therapy alone group. There were statistically significant differences (P < 0.01). Histological analysis using hematoxylin and eosin and Masson’s trichrome staining, along with CD31 immunohistochemistry, showed less lymphocyte infiltration, greater collagen accumulation, and newly formed vessel growth in the high-density nanofat combined with negative pressure wound therapy group, compared with the control group. High-density nanofat grafting combined with negative pressure wound therapy is a safe and effective option, enhancing chronic wound healing.

Keywords: Adipose-derived stem cells, nanofat, negative pressure wound therapy, stromal vascular fraction, wound healing

Introduction

Chronic wounds are characterized as wounds that have not proceeded, within 3 months, through systematic and timely reparation to produce anatomic and functional integrity [1]. Chronic wounds are an important social and economic burden. Incidence of these wounds has continued to increase because of a sharp growth in the aging population and in cardiovascular and metabolic disease, worldwide. It has been estimated that approximately 1-2% of individuals, at any given time, are affected by chronic wounds [2, 3]. Therefore, treatment of chronic wounds is very important and challenging work. However, discovering an optimal therapy for chronic wounds has remained a major challenge for clinicians [4].

Stem cell therapy has been proposed as a treatment for chronic wounds based on the precise physiologic requirements for chronic wound healing, including reepithelization of the wound, angiogenesis, and immunomodulation [5, 6]. Therefore, stem cell therapy has been considered the most promising treatment for chronic wounds [7]. Although bone marrow-derived mesenchymal stem cells (MSCs) were the first stem cells used, clinically, to accelerate wound healing [8, 9], adipose-derived stem cells (ASCs) are superior in many ways, including their ease of harvest, abundance within adipose tissue, and advantages in proangiogenic properties and immunomodulatory effects [10, 11]. Moreover, stromal vascular fraction (SVF) of adipose tissue containing ASCs has been shown to have reparative and regenerating
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Figure 1. Schematic diagram of treatment with high-density nanofat grafting combined with NPWT for chronic wounds. A. Coleman fat after the first centrifugations. B. Nanofat after the second centrifugations: I, low-density nanofat layer; II, high-density nanofat layer; III, aqueous layer. C. High-density nanofat grafting. D. NPWT. NPWT, negative pressure wound therapy.

Negative pressure wound therapy (NPWT), also known as vacuum-assisted closure, micro-deformational wound therapy, or vacuum sealing drainage, is one of the most essential technological advancements in wound care. It has been used as a “bridge” technique for wounds that cannot be immediately closed by either primary intention or plastic surgical techniques [24]. However, the effects of NPWT combined with nanofat grafting on wound healing have not been investigated. Therefore, to increase the concentration of ASCs, this study further processed nanofat through several centrifugations to harvest high-density nanofat. High-density nanofat grafting combined with NPWT was then used to treat patients with chronic wounds, while NPWT alone was used as a control. A comparative study of wound healing rates, 10 days post-surgery, and histological changes was performed. To the best of our knowledge, this is the first study concerning the clinical application of nanofat grafting combined with NPWT for treatment of chronic wounds.

Materials and methods

Ethics approval

Patients with chronic wounds were hospitalized in the Affiliated Hospital of Zunyi Medical College. Before participation, written informed consent was obtained from all patients. This study was approved by the Ethics Committee of the Affiliated Hospital of Zunyi Medical College.

Inclusion and exclusion criteria

Inclusion criteria for the study were: (1) Age between 18 and 80 years; (2) Duration of the
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**Table 1. Summary of patient backgrounds and wound characteristics**

<table>
<thead>
<tr>
<th>Patient number</th>
<th>Age/ Sex</th>
<th>Etiology or risk factors of chronic wound</th>
<th>Wound site</th>
<th>Duration of wound (months)</th>
<th>Original wound size (cm²)</th>
<th>Wound size after 10 days (cm²)</th>
<th>Wound healing rate (%)</th>
<th>Wound treatment</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>42/F</td>
<td>Traumatic infection, hypertension, and obesity</td>
<td>Left leg</td>
<td>3</td>
<td>8.89</td>
<td>4.98</td>
<td>43.98</td>
<td>Nanofat+ NPWT</td>
</tr>
<tr>
<td>2</td>
<td>55/M</td>
<td>Traumatic infection</td>
<td>Right knee</td>
<td>5</td>
<td>4.23</td>
<td>3.49</td>
<td>17.49</td>
<td>Nanofat+ NPWT</td>
</tr>
<tr>
<td>3</td>
<td>46/M</td>
<td>Venous stasis ulcer</td>
<td>Left leg</td>
<td>6</td>
<td>7.86</td>
<td>4.99</td>
<td>36.51</td>
<td>Nanofat+ NPWT</td>
</tr>
<tr>
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<td>Right leg</td>
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<td>21.27</td>
<td>16.05</td>
<td>24.54</td>
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</tr>
<tr>
<td>5</td>
<td>48/M</td>
<td>1. Venous stasis ulcer 2. Great saphenous varices</td>
<td>Left leg</td>
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<td>15.78</td>
<td>11.79</td>
<td>25.29</td>
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</tr>
<tr>
<td>6</td>
<td>51/M</td>
<td>Scar ulcer and traumatic infection</td>
<td>Left leg</td>
<td>8</td>
<td>11.05</td>
<td>8.19</td>
<td>25.88</td>
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<tr>
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<td>Left leg</td>
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<td>9.59</td>
<td>8.11</td>
<td>15.11</td>
<td>Nanofat+ NPWT</td>
</tr>
<tr>
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<td>Right leg</td>
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<td>13.90</td>
<td>10.68</td>
<td>23.21</td>
<td>Nanofat+ NPWT</td>
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<tr>
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</tr>
<tr>
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<tr>
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<td>Right leg</td>
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<td>NPWT</td>
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<tr>
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<td>Left leg</td>
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<tr>
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<tr>
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<td>59/F</td>
<td>Traumatic infection</td>
<td>Right leg</td>
<td>4</td>
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<td>16.17</td>
<td>14.26</td>
<td>NPWT</td>
</tr>
<tr>
<td>15</td>
<td>56/M</td>
<td>Burn and skin tissue infection</td>
<td>Left knee</td>
<td>5</td>
<td>24.78</td>
<td>21.00</td>
<td>15.25</td>
<td>NPWT</td>
</tr>
</tbody>
</table>

F, female; M, male; NPWT, negative pressure wound therapy.
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Figure 2. Wound changes after high-density nanofat grafting combined with NPWT. Patient 1: (A) A chronic left leg wound with an original area of 8.89 cm². (B) Residual area 10 days post-treatment is 4.98 cm². Patient 2: (C) A chronic right knee wound with an original area of 4.23 cm². (D) Residual area 10 days post-treatment is 3.49 cm². NPWT, negative pressure wound therapy.

Figure 3. Wound changes after high-density nanofat grafting combined with NPWT. Patient 7: (A) A diabetic foot wound with an original area of 9.59 cm². (B) Residual area 10 days post-treatment is 8.11 cm². Patient 8: (C) A chronic right leg wound with an original area of 13.90 cm². (D) Residual area 10 days post-treatment is 10.68 cm². NPWT, negative pressure wound therapy.

Harvesting high-density nanofat

Nanofat was prepared according to previously described methods [21]. First, the liquid portion was discarded after centrifuging at 1200 g for 3 minutes. Dry fat was then mechanically emulsified by transferring it between two 10-mL syringes connected by a female-to-female Luer-Lok. After processing, the fat turned into an emulsion and it was filtered using a Nano Transfer filter (Tulip Medical Products, San Diego, CA, USA) to remove connective tissue remnants. Next, the mixture was processed by centrifugation at 2000 g for 3 minutes. This resulted in three layers in the centrifuge tube, low-density nanofat layer, high-density nanofat layer, and a small aqueous layer (Figure 1A-D).

SVF isolation and cell counting

SVF cells were isolated from original nanofat and high-density nanofat. Briefly, the fat was digested (30 minutes in a
shaker, at 37°C) with phosphate buffered saline containing 0.075% collagenase. Mature adipocytes and connective tissue were removed by centrifugation at 800 g for 5 minutes. Cell pellets were then resuspended and filtered through a 100 μm mesh. Finally, the number of SVF cells was counted using an automated cell counter (Countstar, IC1000, Shuangda Biotechnologies Inc, Shanghai, China).

**Surgical intervention of chronic wounds**

Under spinal anesthesia, liposuction of the thighs was performed and high-density nanofat was prepared, as described previously. The wound was debrided and the high-density nanofat was applied. The volume of high-density nanofat applied was based on the area of the wound, approximately 2 mL/cm². Approximately three-fourths of the prepared high-density nanofat was injected directly into the base and edges of the wound. The remaining gel was used to cover the wound as a dressing. Finally, the treated wound was covered with negative pressure drainage. The control group was only treated with negative pressure drainage. Digital photographs of the wounds were taken 10 days postoperatively. Peri-wound tissues were harvested by punch biopsy and fixed in 10% paraformaldehyde for histological analysis.

**Wound healing rates**

On post-surgical day 10, the residual wound area was calculated by the pixel/area ratio using the Adobe Photoshop CS6.0 program (Adobe Systems, San Jose, CA, USA). Finally, wound healing rates were calculated as follows:

\[
\text{Wound healing rate} = \frac{\text{original wound area} - \text{residual wound area}}{\text{original wound area}} \times 100\%.
\]

**Histological analysis**

Peri-wound tissues were fixed in 10% paraformaldehyde, then the tissue samples were embedded in paraffin and cut into approximately 4-mm sections. To observe the tissue morphology, the sections were processed and stained with hematoxylin and eosin (H&E). To observe collagen deposition, the sections were stained.

**Figure 4.** Wound changes after treatment with NPWT alone. Patient 10: (A) A chronic left heel wound with an original area of 11.64 cm². (B) Residual area 10 days post-treatment is 11.07 cm². Patient 13: (C) A chronic right hand wound with an original area of 5.55 cm². (D) Residual area 10 days post-treatment is 5.10 cm². (E) Quantification of the wound area of the different treatment groups. **P < 0.01. NPWT, negative pressure wound therapy.
with Masson’s trichrome. Relative collagen expression was calculated using Image J (National Institute of Mental Health, Bethesda, MD, USA). Neovascularization was evaluated after CD31 immunostaining and the number of new blood vessels per 5 high-power fields was counted.

Statistical analysis

Results are expressed as mean ± standard deviation. Data were analyzed with independent sample t-tests using SPSS version 19.0 (IBM Corp., Armonk, NY, USA). P < 0.05 indicates statistical significance.

Results

Cellular components of high-density nanofat

SVF cell numbers in each sample suggested that SVF cell density increased sharply after mechanical processing (8.2 × 10^4 ± 1.4 × 10^4 cells/mL in original nanofat and 2.2 × 10^5 ± 0.3 × 10^5 cells/mL in high-density nanofat).

Wound healing rates

Fifteen patients admitted to the Affiliated Hospital of Zunyi Medical College, between January 2016 and October 2017, were enrolled and evaluated (11 men and 4 women, aged 42-80 years). Wound types included 6 venous stasis ulcers, 6 traumatic infections, 1 scar ulcer, 1 diabetic foot, and 1 burn wound (Table 1). Although these complex chronic wounds were treated with different interventions after transplantation of autologous high-density nanofat, all patients achieved good outcomes. Average wound healing rate for the 8 patients that received high-density nanofat combined with NPWT was 26.50% ± 9.51%. The rate was 12.02% ± 4.20% for the 7 patients receiving NPWT alone. Differences were statistically significant (P < 0.01) (Figures 2A-D, 3A-D, 4A-E). Although nanofat was harvested from the thighs, ipsilaterally or bilaterally, there were no donor site complications.

Histological analysis

H&E staining of paraffin specimens from the peri-wound tissue revealed that lymphocyte infiltration of the deep dermal layers was decreased, indicating a reduction in the lymphocyte-mediated chronic inflammatory response following high-density nanofat administration 10 days after treatment (Figure 5A-D). Tissues stained with Masson’s trichrome stain were examined for collagen deposition, revealing greater collagen accumulation in the high-density nanofat combined with NPWT group than in the NPWT alone group (P < 0.001). Collagen was obviously thicker in the nanofat combined with NPWT group than in the NPWT alone group (Figure 6A-E). Immuno histoche-

mistry with CD31 indicated more newly formed vessels in the nanofat combined with NPWT group than in the NPWT alone group (P < 0.001) (Figure 7A-E).

Discussion

Although moderate beneficial effects of nanofat grafting or NPWT for acceleration of wound healing have been reported [22, 25], the effects of nanofat grafting combined with NPWT on wound healing have not been investigated. The present study found that high-density nanofat grafting combined with NPWT can markedly enhance wound healing rates, compared to NPWT alone. In addition, histological analysis indicated that nanofat grafting combined with NPWT accelerates wound healing by reducing the infiltration of inflammatory cells, increasing collagen deposition, and promoting neovascularization. Collectively, present results suggest that there are synergistic effects between nanofat and NPWT that promote accelerated wound healing.

Currently, NPWT is one of the most widespread techniques used to treat chronic wounds [25]. Possible mechanisms of NPWT in wound care include the stimulation of granulation tissue formation, decrease in matrix metalloproteinases, stimulation of angiogenesis, and less edema formation, leading to improved blood flow and reduced bacterial colonization [25-27]. However, chronic wounds require multiple treatments because of the dynamic and irregular wound environment [9]. Hence, it is improbable that one type of treatment alone will result in complete wound healing [9]. A recent large randomized trial suggested that the most important function of NPWT is to promote granulation tissue and generate a “curative” environment inside a wound rather than specifically to achieve wound closure [28]. Similarly, present results demonstrated that the wound healing rate at 10 days after treatment was only 12.02% ± 4.20% in the NPWT alone group. However, this treatment resulted in better granulation tissue formation. Similarly, Langer et al. [25] reported that there were statistically significant differences in the volume of the wounds before and after the application of NPWT. Average reduction in wound size per week was 17.44% in chronic
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lower limb wounds in their 34 patients. Minor differences in wound healing rates may be related to the duration of chronic wounds, severity of the condition, and negative pressure suction. However, in contrast, the average wound healing rate of high-density nanofat combined with NPWT group was 26.50% ± 9.51%, considerably higher than that of the NPWT alone group. Differences in wound healing rates may be due to the transplantation of high-density nanofat.

Traditional nanofat includes water, oil droplets, SVF, and extracellular matrix. Concentrations of ASCs within nanofat are very low, only $1 \times 10^3$/mL. This is much lower than cell concentrations required for stem cell therapy. Therefore, traditional nanofat is further processed and purified through a purely physical method, resulting in high-density nanofat whose volume is approximately 30% of the initial nanofat volume. As a result, concentrations of ASCs within the high-density nanofat are effectively improved, increasing the clinical benefits. Present data showed that the number of SVF cells in original nanofat was $8.2 \times 10^4 \pm 1.4 \times 10^4$ cells/mL while that in high-density nanofat was $2.2 \times 10^5 \pm 0.3 \times 10^5$ cells/mL. It has been established that ASCs have the potential to promote angiogenesis and secrete a significant amount of growth factors, showing immunomodulatory effects [29]. In the present study, H&E staining revealed that the lymphocyte infiltration in the dermal layers was obviously decreased after injections of high-density nanofat. Results confirm the immunomodulatory effects of ASCs on lymphocyte-mediated chronic inflammatory response. Additionally, collagen synthesis and neovascularization is essential for wound healing [30]. Histological analysis demonstrated that collagen accumulation and newly formed vessels were higher in the high-density nanofat combined with NPWT group than in the NPWT alone group. Results suggest that there could be synergistic effects between high-density nanofat and NPWT that promote accelerated wound healing. Similarly, Sahin et

![Figure 7. Immunohistochemistry analysis of the wound area with CD31. A. Rare neovascularization before NPWT alone. B. Rare neovascularization before high-density nanofat grafting combined with NPWT. C. Neovascularization is increased after NPWT alone. D. The most neovascularization is seen after high-density nanofat grafting combined with NPWT. E. Relative quantification of neovascularization. ***P < 0.001. NPWT, negative pressure wound therapy.](image-url)
al. [31] reported that NPWT combined with MSCs improved the neovascularization of an acellular dermal matrix (ADM) application to Sprague-Dawley rats with wounds. Results revealed that the application of NPWT with MSCs to wounds significantly accelerated the neovascularization of ADM, more than NPWT or MSCs alone, suggesting that the combination of NPWT with MSCs has synergistic effects on angiogenesis.

There were limitations to this study, however. The information presented in this study was limited by its small sample size, although the patients obtained satisfactory outcomes. In addition, because of ethical issues, single injections of high-density nanofat were not included in the control group. Moreover, optimal volumes, transplant timing, and number of transplants of high-density nanofat requires further study. Once a standard protocol has been identified, a large multicenter clinical controlled trial should be performed. However, the present study indicates that high-density nanofat transplantation is a valuable treatment for chronic wounds.

**Conclusion**

High-density nanofat grafting combined with NPWT can accelerate wound healing by reducing the infiltration of inflammatory cells, increasing collagen deposition, and promoting neovascularization. There may be synergistic effects between nanofat and NPWT on wound healing. This combination can be an effective and safe option for treatment of chronic wounds.

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**Disclosure of conflict of interest**

None.

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