Original Article

Protective effects of wogonoside against β-amyloid-induced neurotoxicity and neuroinflammation

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Abstract: Accumulation of beta-amyloid (Aβ) is one of the most important features of Alzheimer's disease (AD), which plays a critical role in AD-related neurotoxicity, neuroinflammation, oxidative stress and neuronal cell death. In the current study, we were aiming to investigate the effects of wogonoside on $Aβ_{1.42}$ -induced neurotoxicity and neuroinflammation. We found in HT22 cells, wogonoside treatment significantly ameliorated $Aβ_{1.42}$ -induced cell death, cell apoptosis and suppressed the release of lactate dehydrogenase (LDH). Moreover, we observed that in BV-2 cells, wogonoside treatment inhibited pro-inflammatory cytokines production and oxidative stress induced by $Aβ_{1.42}$. Furthermore, we found that wogonoside treatment significantly increased Bcl-2/Bax ratio, suppressed Akt/NF-κB pathway but activated Nrf2/HO-1 pathway. In conclusion, our study demonstrated that wogonoside may exert anti-apoptotic, anti-inflammatory and anti-oxidative effects against Aβ-induced neurotoxicity and neuroinflammation, and suggested wogonoside as potential drug candidate for AD.

Keywords: Beta-amyloid, Alzheimer's disease, wogonoside, Akt/NF-кВ, Nrf2/HO-1

Introduction

Alzheimer's disease (AD) is the most common age-related neurodegenerative disease in the elderly [1, 2]. The main pathological features of AD are the extracellular formation of beta-amyloid (A β) plaques and the intracellular formation of neurofibrillary tangles [3]. Although the exact cause and related mechanisms of AD still remain largely unknown, many evidences show that neuronal apoptosis, mitochondria dysfunction and oxidative stress played critical roles in the pathogenesis of AD [4-7].

Microglia-mediated neuroinflammation, characterized by excessive microglia activation and overproduction of pro-inflammatory cytokines and chemokines, is also an important component of AD, which starts as a defense mechanism against the A β deposition in the brain, but can also lead to neurodegeneration [8]. Excessive activation of microglia will not only release inflammatory cytokines, but will also synthesize and release some cytotoxic factors, such as nitric oxide and reactive oxygen species, leading to significant neuronal cell damage [9]. This indicates that inhibition of microg-

lia-mediated inflammatory responses can play a potential therapeutic role in the treatment of AD.

More than 40 flavonoids derivatives have been extracted and identified from the roots of Scutellaria baicalensis Georgi [10]. Among them, the most abundant content is baicalin and wogonoside [11]. Both baicalin and wogonoside were found to exhibit anti-tumor, anti-oxidative and anti-inflammatory activities [12-19]. Recently, it has been reported that baicalin has protective effects on AB-induced cognitive impairment, oxidative stress and neuronal apoptosis in rat [20, 21], as well as the inhibitory effects on AB-induced microglial cell activation [22], which suggested the protective role of baicalin in the pathogenesis of AD. However, the effect of wogonoside on AD, especially on Aβ₁₋₄₂-induced neurotoxicity and neuroinflammation, has not been reported.

In this study, we were aiming to investigate the role of wogonoside in A β -induced neurotoxicity and neuroinflammation in cultured HT22 hippocampal cells and BV-2 microglial cells. We found that wogonoside treatment significantly in-

creased HT22 cell viability, inhibited the release of LDH and suppressed cell apoptosis after $\mathsf{A}\beta_{1.42}$ challenge. Furthermore, we observed that treatment of wogonoside increased the expression of Bcl-2 and decreased Bax protein level, leading to the increased level of Bcl-2/Bax ratio. Moreover, we found wogonoside treatment decreased the expression of proinflammatory cytokines, ameliorated $\mathsf{A}\beta_{1.42}$ -induced oxidative stress in $\mathsf{A}\beta_{1.42}$ -treated BV-2 cells, which may relate to the suppressed Akt/NF-kB pathway and activated Nrf2/HO-1 pathway modulated by wogonoside.

Material and methods

Cells and materials

HT22 hippocampal cells and BV-2 microglial cells were obtained from American Type Culture Collection (Manassas, VA). A β_{1-42} was purchased from American Peptide Company (Sunnyvale, CA, USA). Wogonoside was purchased from Langze Pharmaceutical Company (Nanjing, China). Antibodies used for β-actin (#3700), Nrf2 (#12721), HO-1(#70081), Bax (#2772), Bcl-2 (#2764), p-Akt (#4060), Akt (#4685), p-P65 (#3033), P65 (#8242) were all obtained from Cell Signaling Technology.

Flow cytometry and caspase assay

After pre-incubation with wogonoside followed by $A\beta_{1.42}$ treatment, HT22 cells were collected and suspended in 100 µL binding buffer. 5 µL of Annexin V-PE and 5 µL of 7-amino-actinomycin D (7-AAD) were added and the mixture was incubated in the dark for 15 min. The caspase-3 activity was examined by commercial detection kit (ab39401, Abcam) according to the manufacturer's protocol.

Cell viability assay and Lactate dehydrogenase (LDH) activity assay

A commercial LDH diagnostic kit (STANBIO Laboratory, Boerne, TX, USA) was used to measure LDH activity according to the manufacturer's instructions. The procedure of cell viability assay was as follow: After pre-incubation with wogonoside followed by A β_{1-42} treatment, 20 μ L MTT was added to each well and incubated at 37°C. Four hours later, the supernatants were aspirated off and 130 μ L of DMSO was added to. SpectraMax Plus384 Microplate Reader

(Molecular Devices, USA) was used to detect the value of optical density.

ELISA and gRT-PCR

TNF- α and IL-6 ELISA kits (R&D Systems, Minneapolis, MN, USA) were used to measure the protein levels of TNF-α and IL-6 in cell-free supernatants. Total RNA was extracted by TRIzol reagent (Invitrogen). Synthesis of cDNA was performed with M-MLV First Strand Kit (Taraka, China). Real-time PCR was implemented with GoTag gRT-PCR Master Mix (Promega, USA). GAPDH was used as the internal control, and the 2^- $\Delta\Delta$ CT method was used to analyze the results. Primer sequences: TNF-α: 5'-AGG GCC ATT CCT ACT CCC AT-3' (F), 5'-TGT AGC CCC GGA TAC ACA GA-3' (R); IL-6: 5'-CAC TTC ACA AGT CGG AGG CT-3' (F). 5'-TCT GAC AGT GCA TCA TCG CT-3' (R); GAPDH: 5'-GGA GAG TGT TTC CTC GTC CC-3' (F), 5'-ACT GTG CCG TTG AAT TTG CC-3'(R).

Measurement of intracellular MDA and activities of SOD and CAT

MDA level was measured with a commercial kit (Nanjing Jiancheng Biochemistry Co., Nanjing, China) according to the manufacturer's instructions. The activities of SOD and CAT were measured using colorimetric assay kits (Abcam, USA) according to the protocols provided by the manufacturer.

Western blot analysis

Cell were collected and lysed with RIPA Lysis (Thermo Fisher scientific) with a protease inhibitor cocktail tablet (Roche diagnostic, Basel, Switzerland). Western blot analysis was performed as described [23].

Statistical analysis

All the data were reported as mean \pm S.D. and statistical differences were analyzed by one-way ANOVA followed by Turkey test. P<0.05 was considered statistically significant.

Results

Wogonoside attenuated $A\beta_{1-42}$ -induced cytotoxicity

To investigate the effect of wogonoside on $A\beta_{\text{1-42}}\text{-induced}$ neurotoxicity, we firstly exam-

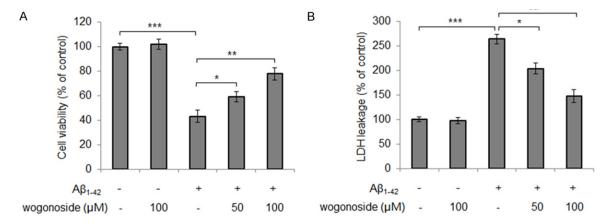


Figure 1. Wogonoside attenuated $A\beta_{1.42}$ -induced cytotoxicity. HT22 cells were incubated without or with wogonoside (50, 100 μ M) for 4.0 h, followed by incubation with $A\beta_{1.42}$ for another 24 h. A. Cell viability was determined with the MTT assay. B. The release of LDH into extracellular surrounding was measured with LDH assay. Results are shown as the mean \pm S.D. and represent three independent experiments. *, P<0.05; **, P<0.01; ***, P<0.001.

ined the role of wogonoside in cell viability in $A\beta_{1.42}$ -treated HT22 cells. As shown in **Figure 1A**, treatment of HT22 cells with $A\beta_{1.42}$ significantly decreased the level of cell viability, but when the cells were pretreated with wogonoside followed by exposure to $A\beta_{1.42}$, the level of cell viability was increased in a wogonoside dose-dependent manner. Next, we determined the level of LDH release, which is also an indicator of cell injury, from $A\beta_{1.42}$ -treated HT22 cells. As shown in **Figure 1B**, the level of LDH release was significantly improved by $A\beta_{1.42}$ treatment. Conversely, in the presence of wogonoside, LDH activity was obviously reduced compared to the control group.

Wogonoside suppressed $A\beta_{1-42}$ -induced cell apoptosis

Apoptotic cell death plays a vital role in the pathogenesis of AD. Therefore we used flow cytometry analysis to detect the effect of wogonoside on $A\beta_{1-42}$ -induced cell apoptosis. Consistently with previous studies, we found $A\beta_{1-42}$ treatment indeed promoted cell apoptosis in HT22 cells. However, pre-incubation with wogonoside resulted in a significant decrease in the apoptotic rate (Figure 2A and 2B). Furthermore, we examined the activity of caspase-3 and we found treatment of wogonoside decreased caspase-3 activity in a dose-dependent manner (Figure 2C). As shown in Figure 2D, we observed that wogonoside treatment significantly increased the protein level of Bcl-2 and inhibited the expression of Bax, leading to the increased Bcl-2/Bax ratio (Figure 2E) in $A\beta_{1.42}$ -treated HT22 cells, which indicated that wogonoside could regulate $A\beta_{1-42}$ -induced HT22 cell apoptosis by modulate the expression level of Bax and Bcl-2.

Wogonoside inhibited $A\beta_{1.42}$ -induced pro-inflammatory cytokines production in microglial cells

Wogonoside has been reported to play anti-inflammatory roles in diverse disease models [12, 15]. Therefore, we examined the effect of wogonoside on A β_{1-42} -induced inflammation in BV-2 microglial cells. We found the mRNA levels of pro-inflammatory cytokines such as TNF- α and IL-6 were both up-regulated by A β_{1-42} challenge, but pre-treatment with wogonoside significantly decreased the expression of these cytokines in a dose-dependent manner (**Figure 3A**). Consistently, the secreted protein levels of TNF- α and IL-6 in wogonoside-treated groups were also decreased compared to the control group (**Figure 3B**).

Wogonoside attenuated $A\beta_{1-42}$ -induced oxidative stress in microglial cells

Furthermore, we examined the effect of wogonoside on $A\beta_{1-42}$ -induced oxidative stress. As shown in **Figure 4**, oxidative stress was assessed by measuring the level of malondialdehyde (MDA) (**Figure 4A**), and the activities of antioxidant enzymes including superoxide dismutase (SOD) (**Figure 4B**) and catalase (CAT) (**Figure 4C**). We found pre-treatment of wogonoside significantly reduced the produc-

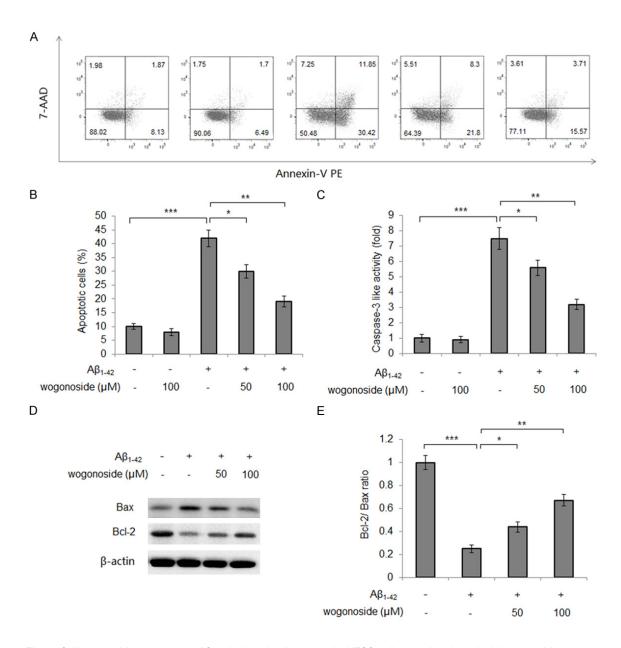


Figure 2. Wogonoside suppressed $A\beta_{1.42}$ -induced cell apoptosis. HT22 cells were incubated without or with wogonoside (50, 100 μ M) for 4.0 h, followed by incubation with $A\beta_{1.42}$ for another 24 h. (A and B) The percentage of apoptotic cells was measured by flow cytometry analysis. (C) Caspase-3 activity was detected with a fluorometric assay. (D) Protein levels of Bax and Bcl-2 were examined by western blot. (E) Quantification of protein levels of Bax and Bcl-2 in (D). Results are shown as the mean \pm S.D. and represent three independent experiments. *, P<0.05; **, P<0.01; ***, P<0.001.

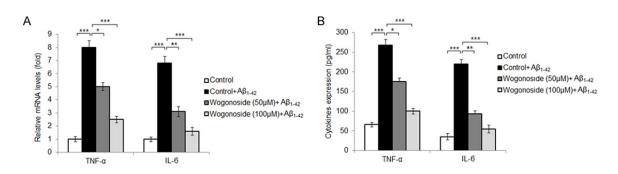


Figure 3. Wogonoside inhibited $Aβ_{1.42}$ -induced pro-inflammatory cytokines production in microglial cells. BV-2 cells were incubated without or with wogonoside (50, 100 μM) for 4.0 h, followed by incubation with $Aβ_{1.42}$ for another 24 h. A. mRNA levels of TNF-α and IL-6 were examined by qRT-PCR. B. Protein levels of TNF-α and IL-6 were measured by ELISA. Results are shown as the mean \pm S.D. and represent three independent experiments. *, P<0.05; **, P<0.01; ***, P<0.001.

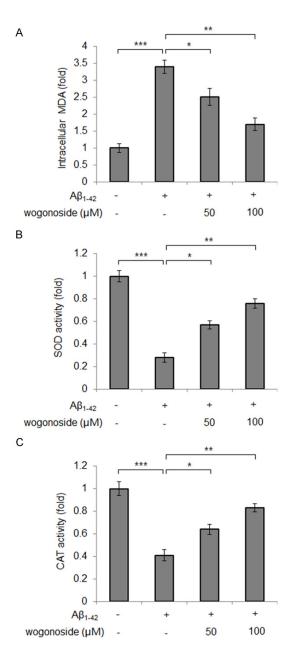


Figure 4. Wogonoside attenuated A $\beta_{1.42}$ -induced oxidative stress in microglial cells. BV-2 cells were incubated without or with wogonoside (50, 100 μ M) for 4.0 h, followed by incubation with A $\beta_{1.42}$ for another 24 h. Intracellular MDA level (A), SOD activity (B) and CAT activity (C) were detected. *, P<0.05; **, P<0.01; ***, P<0.001.

tion of the oxidative marker MDA in $A\beta_{1-42}$ -treated BV-2 cells, and increased the activities

of antioxidant enzymes such as SOD and CAT in a dose-dependent manner.

Wogonoside suppressed Akt/NF- κ B pathway and activated Nrf2/HO-1 pathway in A β_{1-42} -treated microglial cells

Previous studies revealed that wogonoside plays an inhibitory role in Akt/NF-kB pathway [12, 15]. In order to investigate how wogonoside regulates the pro-inflammatory cytokines production and oxidative stress, we examined the protein levels of p-Akt, Akt, p-P65 and P65 in $A\beta_{1.42}$ -treated microglial cells (**Figure 5A**). We found that the expression of p-Akt and p-P65 was significantly reduced in wogonoside-administrated BV-2 cells in a dose-dependent manner (Figure 5B and 5C). It has been reported that Nrf2/HO-1 pathway plays critical roles in controlling antioxidant responses and inhibition of NF-kB signaling in microglia cells [24, 25], therefore we also examined the effect of wogonoside on Nrf2 and HO-1 expression. We found the protein levels of Nrf2 and HO-1 were both decreased in BV-2 cells after $A\beta_{1-42}$ treatment (Figure 5A). Most importantly, we found wogonoside could up-regulate the expression of Nrf2 and HO-1 in a dose-dependent manner (**Figure 5D** and **5E**) in $A\beta_{1-42}$ -treated BV-2 microglial cells.

Discussion

In the current study, in order to investigate the function of wogonoside in AD, we examined the effect of wogonoside on A β_{1-42} -induced neurotoxicity and neuroinflammation in cultured HT22 hippocampal cells and BV-2 microglial cells. To the best of our knowledge, this is the first article to demonstrate the protective role of wogonoside against A β_{1-42} -induced nervous system disease.

Alzheimer's disease is the most common neurodegenerative disorder in the world [26]. This disease is characterized by synaptic impairment [27], neurotrophin and neurotransmitter imbalance, mitochondrial dysfunction, oxidative stress, intracellular calcium increase and

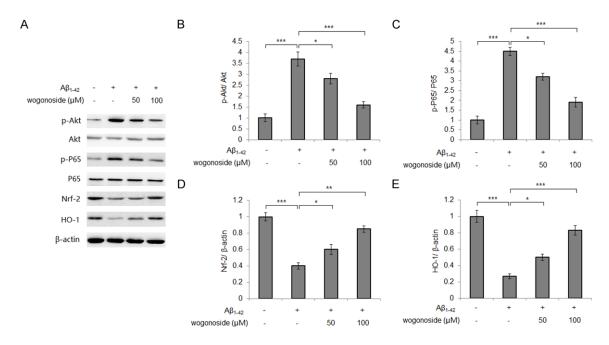


Figure 5. Wogonoside suppressed Akt/NF-κB pathway and activated Nrf2/HO-1 pathway in $Aβ_{1.42}$ -treated microglial cells. BV-2 cells were incubated without or with wogonoside (50, 100 μM) for 4.0 h, followed by incubation with $Aβ_{1.42}$ for another 24 h. A. Protein levels of p-Akt, Akt, p-P65, P65, Nrf-2 and HO-1 were examined by western blot. B-E. Quantification of protein levels of p-Akt, p-P65, Nrf-2 and HO-1 in (A). *, P<0.05; ***, P<0.01; ****, P<0.001.

cell cycle failure [28]. The most severe changes are in hippocampus, as well as in cortical and subcortical regions [29], which are associated with the AD memory deficits [30]. Abnormal cleavage of amyloid precursor protein results in the accumulation of AB in the brain, and leads to the pathological manifestations of AD [31]. It has been well studied that treatment with or intracerebral injection of $A\beta_{1-42}$ significantly induced neurotoxicity and AD related cognitive and cerebral changes in vivo and in vitro [32, 33]. Similar with their findings, in the current study, we found stimulated with $A\beta_{1,42}$ indeed decreased the cell viability of HT22 cells, increased the release of LDH and promoted cell apoptosis. However, we found pre-treatment with wogonoside significantly reversed these phenomenon induced by $A\beta_{1.42}$, and wogonoside treatment increased Bcl-2/Bax ratio which indicated that wogonoside could protect HT22 cells from $A\beta_{1-42}$ -induced cell apoptosis and neurotoxicity.

Oxidative stress has been found to be implicated in the neurotoxicity of $A\beta$ and in the pathogenesis of AD, and there is growing evidence that oxidative stress is closely related to the inflammatory response of microglia [34, 35]. Consistently, we found $A\beta_{1.42}$ treatment significantly increased the level of MDA and sup-

pressed the activities of SOD and CAT in BV-2 cells. Whereas after pre-incubation with wogonoside, MDA level in BV-2 cells was greatly decreased, as well as the increased activities of SOD and CAT. Similar with oxidative stress, neuroinflammation, characterized by excessive glial activation and overproduction of proinflammatory cytokines and chemokines, was also found to play critical roles in the pathogenesis of neurodegeneration in AD and related neurodegenerative disorders [36]. In the current study, we observed that both mRNA and protein levels of pro-inflammatory cytokines such as TNF- α and IL-6 induced by A $\beta_{1,42}$ were significantly decreased in the presence of wogonoside. Our findings suggested that wogonoside played anti-apoptotic, anti-oxidative and anti-inflammatory roles in the protection against $A\beta_{1-42}$ stimulation.

Akt/NF-κB signaling has represented a paradigm for signal transduction and pro-inflammatory cytokines production implicated in numerous diseases including AD [37-39]. Previous studies also reported that wogonoside could inhibit Akt/NF-κB signaling activation in diverse diseases [12, 15]. Consistently, in the present research, we found wogonoside treatment greatly inhibited Akt/NF-κB pathway activation through suppressing the phosphorylation of Akt

and P65, and this inhibition may lead to the decreased production of pro-inflammatory cytokines. Nrf2 is crucial for the regulation of antioxidant genes, including H0-1 and NQ01, and it has been reported that the Nrf2/H0-1 signaling pathway plays an important role in the action of neuroprotectant [40, 41]. In this study, for the first time, we found wogonoside induced H0-1 expression and increased Nrf2 levels in a dose-dependent manner in $A\beta_{1-42}$ -stimulated BV-2 cells.

In conclusion, these results indicated that wo-gonoside could increase Bcl-2/Bax ratio, inhibit Akt/NF- κ B signaling and activate Nrf2/HO-1 pathway, therefore played anti-apoptotic, anti-oxidative and anti-inflammatory roles to protect HT22 hippocampal cells and BV-2 microglial cells against A β_{1-42} -induced neurotoxicity and neuroinflammation.

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Disclosure of conflict of interest

None.

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References

- [1] Yamin G, Ono K, Inayathullah M and Teplow DB. Amyloid beta-protein assembly as a therapeutic target of Alzheimer's disease. Curr Pharm Des 2008; 14: 3231-3246.
- [2] van der Flier WM and Scheltens P. Epidemiology and risk factors of dementia. J Neurol Neurosurg Psychiatry 2005; 76 Suppl 5: v2-7.
- [3] Braak H and Braak E. Alzheimer's disease: striatal amyloid deposits and neurofibrillary changes. J Neuropathol Exp Neurol 1990; 49: 215-224.
- [4] Billings LM, Oddo S, Green KN, McGaugh JL and LaFerla FM. Intraneuronal Abeta causes the onset of early Alzheimer's disease-related cognitive deficits in transgenic mice. Neuron 2005; 45: 675-688.
- [5] Walsh DM and Selkoe DJ. Oligomers on the brain: the emerging role of soluble protein aggregates in neurodegeneration. Protein Pept Lett 2004; 11: 213-228.

- [6] Choi H, Park HH, Koh SH, Choi NY, Yu HJ, Park J, Lee YJ and Lee KY. Coenzyme Q10 protects against amyloid beta-induced neuronal cell death by inhibiting oxidative stress and activating the P13K pathway. Neurotoxicology 2012; 33: 85-90.
- [7] Shen WX, Chen JH, Lu JH, Peng YP and Qiu YH. TGF-beta1 protection against Abeta1-42-induced neuroinflammation and neurodegeneration in rats. Int J Mol Sci 2014; 15: 22092-22108.
- [8] Giovannini MG, Scali C, Prosperi C, Bellucci A, Vannucchi MG, Rosi S, Pepeu G and Casamenti F. Beta-amyloid-induced inflammation and cholinergic hypofunction in the rat brain in vivo: involvement of the p38MAPK pathway. Neurobiol Dis 2002; 11: 257-274.
- [9] Mosher KI and Wyss-Coray T. Microglial dysfunction in brain aging and Alzheimer's disease. Biochem Pharmacol 2014; 88: 594-604.
- [10] Li C, Lin G and Zuo Z. Pharmacological effects and pharmacokinetics properties of Radix Scutellariae and its bioactive flavones. Biopharm Drug Dispos 2011; 32: 427-445.
- [11] Lee KJ, Jung PM, Oh YC, Song NY, Kim T and Ma JY. Extraction and bioactivity analysis of major flavones compounds from scutellaria baicalensis using in vitro assay and online screening HPLC-ABTS system. J Anal Methods Chem 2014; 2014: 563702.
- [12] Sun Y, Zhao Y, Wang X, Zhao L, Li W, Ding Y, Kong L, Guo Q and Lu N. Wogonoside prevents colitis-associated colorectal carcinogenesis and colon cancer progression in inflammationrelated microenvironment via inhibiting NFkappaB activation through PI3K/Akt pathway. Oncotarget 2016; 7: 34300-34315.
- [13] Chen Y, Hui H, Yang H, Zhao K, Qin Y, Gu C, Wang X, Lu N and Guo Q. Wogonoside induces cell cycle arrest and differentiation by affecting expression and subcellular localization of PLSCR1 in AML cells. Blood 2013; 121: 3682-3691.
- [14] Chen Y, Lu N, Ling Y, Gao Y, Wang L, Sun Y, Qi Q, Feng F, Liu W, Liu W, You Q and Guo Q. Wogonoside inhibits lipopolysaccharide-induced angiogenesis in vitro and in vivo via tolllike receptor 4 signal transduction. Toxicology 2009; 259: 10-17.
- [15] Sun Y, Zhao Y, Yao J, Zhao L, Wu Z, Wang Y, Pan D, Miao H, Guo Q and Lu N. Wogonoside protects against dextran sulfate sodium-induced experimental colitis in mice by inhibiting NF-kappaB and NLRP3 inflammasome activation. Biochem Pharmacol 2015; 94: 142-154.
- [16] Shi H, Ren K, Lv B, Zhang W, Zhao Y, Tan RX and Li E. Baicalin from Scutellaria baicalensis blocks respiratory syncytial virus (RSV) infection and reduces inflammatory cell infiltration

- and lung injury in mice. Sci Rep 2016; 6: 35851.
- [17] Lin M, Li L, Zhang Y, Zheng L, Xu M, Rong R and Zhu T. Baicalin ameliorates H2O2 induced cytotoxicity in HK-2 cells through the inhibition of ER stress and the activation of Nrf2 signaling. Int J Mol Sci 2014; 15: 12507-12522.
- [18] Chen H, Xu Y, Wang J, Zhao W and Ruan H. Baicalin ameliorates isoproterenol-induced acute myocardial infarction through iNOS, inflammation and oxidative stress in rat. Int J Clin Exp Pathol 2015; 8: 10139-10147.
- [19] Yu Y, Pei M and Li L. Baicalin induces apoptosis in hepatic cancer cells in vitro and suppresses tumor growth in vivo. Int J Clin Exp Med 2015; 8: 8958-8967.
- [20] Chen C, Li X, Gao P, Tu Y, Zhao M, Li J, Zhang S and Liang H. Baicalin attenuates alzheimerlike pathological changes and memory deficits induced by amyloid beta1-42 protein. Metab Brain Dis 2015; 30: 537-544.
- [21] Ding H, Wang H, Zhao Y, Sun D and Zhai X. Protective effects of baicalin on abeta(1)(-)(4) (2)-Induced learning and memory deficit, oxidative stress, and apoptosis in rat. Cell Mol Neurobiol 2015; 35: 623-632.
- [22] Xiong J, Wang C, Chen H, Hu Y, Tian L, Pan J and Geng M. Abeta-induced microglial cell activation is inhibited by baicalin through the JAK2/STAT3 signaling pathway. Int J Neurosci 2014; 124: 609-620.
- [23] Sakae N, Liu CC, Shinohara M, Frisch-Daiello J, Ma L, Yamazaki Y, Tachibana M, Younkin L, Kurti A, Carrasquillo MM, Zou F, Sevlever D, Bisceglio G, Gan M, Fol R, Knight P, Wang M, Han X, Fryer JD, Fitzgerald ML, Ohyagi Y, Younkin SG, Bu G and Kanekiyo T. ABCA7 deficiency accelerates Amyloid-beta generation and Alzheimer's neuronal pathology. J Neurosci 2016; 36: 3848-3859.
- [24] Park SY, Jin ML, Wang Z, Park G and Choi YW. 2,3,4',5-tetrahydroxystilbene-2-0-beta-d-glucoside exerts anti-inflammatory effects on lipopolysaccharide-stimulated microglia by inhibiting NF-kappaB and activating AMPK/Nrf2 pathways. Food Chem Toxicol 2016; 97: 159-167.
- [25] Cheng-Chung Wei J, Huang HC, Chen WJ, Huang CN, Peng CH and Lin CL. Epigallocatechin gallate attenuates amyloid beta-induced inflammation and neurotoxicity in EOC 13.31 microglia. Eur J Pharmacol 2016; 770: 16-24.
- [26] Shi JM, He X, Lian HJ, Yuan DY, Hu QY, Sun ZQ, Li YS and Zeng YW. Tibetan medicine "RNSP" in treatment of Alzheimer disease. Int J Clin Exp Med 2015; 8: 19874-19880.
- [27] Canas PM, Porciuncula LO, Cunha GM, Silva CG, Machado NJ, Oliveira JM, Oliveira CR and

- Cunha RA. Adenosine A2A receptor blockade prevents synaptotoxicity and memory dysfunction caused by beta-amyloid peptides via p38 mitogen-activated protein kinase pathway. J Neurosci 2009; 29: 14741-14751.
- [28] Querfurth HW and LaFerla FM. Alzheimer's disease. N Engl J Med 2010; 362: 329-344.
- [29] Arnold SE, Hyman BT, Flory J, Damasio AR and Van Hoesen GW. The topographical and neuro-anatomical distribution of neurofibrillary tangles and neuritic plaques in the cerebral cortex of patients with Alzheimer's disease. Cereb Cortex 1991; 1: 103-116.
- [30] Wirth M, Madison CM, Rabinovici GD, Oh H, Landau SM and Jagust WJ. Alzheimer's disease neurodegenerative biomarkers are associated with decreased cognitive function but not beta-amyloid in cognitively normal older individuals. J Neurosci 2013; 33: 5553-5563.
- [31] Murphy MP and LeVine H 3rd. Alzheimer's disease and the amyloid-beta peptide. J Alzheimers Dis 2010; 19: 311-323.
- [32] Xuan A, Long D, Li J, Ji W, Zhang M, Hong L and Liu J. Hydrogen sulfide attenuates spatial memory impairment and hippocampal neuroinflammation in beta-amyloid rat model of Alzheimer's disease. J Neuroinflammation 2012; 9: 202.
- [33] Chang W and Teng J. Beta-asarone prevents Abeta25-35-induced inflammatory responses and autophagy in SH-SY5Y cells: down expression Beclin-1, LC3B and up expression Bcl-2. Int J Clin Exp Med 2015; 8: 20658-20663.
- [34] Behl C, Davis JB, Lesley R and Schubert D. Hydrogen peroxide mediates amyloid beta protein toxicity. Cell 1994; 77: 817-827.
- [35] Surace MJ and Block ML. Targeting microgliamediated neurotoxicity: the potential of NOX2 inhibitors. Cell Mol Life Sci 2012; 69: 2409-2427
- [36] Niranjan R. Molecular basis of etiological implications in Alzheimer's disease: focus on neuroinflammation. Mol Neurobiol 2013; 48: 412-428.
- [37] Boissiere F, Hunot S, Faucheux B, Duyckaerts C, Hauw JJ, Agid Y and Hirsch EC. Nuclear translocation of NF-kappaB in cholinergic neurons of patients with Alzheimer's disease. Neuroreport 1997; 8: 2849-2852.
- [38] Srinivasan M and Lahiri DK. Significance of NFkappaB as a pivotal therapeutic target in the neurodegenerative pathologies of Alzheimer's disease and multiple sclerosis. Expert Opin Ther Targets 2015; 19: 471-487.
- [39] Ito S, Sawada M, Haneda M, Fujii S, Oh-Hashi K, Kiuchi K, Takahashi M and Isobe K. Amyloidbeta peptides induce cell proliferation and macrophage colony-stimulating factor expres-

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- sion via the PI3-kinase/Akt pathway in cultured Ra2 microglial cells. FEBS Lett 2005; 579: 1995-2000.
- [40] Yu L, Wang S, Chen X, Yang H, Li X, Xu Y and Zhu X. Orientin alleviates cognitive deficits and oxidative stress in Abeta1-42-induced mouse model of Alzheimer's disease. Life Sci 2015; 121: 104-109.
- [41] Kwon SH, Ma SX, Hwang JY, Lee SY and Jang CG. Involvement of the Nrf2/H0-1 signaling pathway in sulfuretin-induced protection against amyloid beta25-35 neurotoxicity. Neuroscience 2015; 304: 14-28.