Association between aldosterone synthase (CYP11B2) -344C/T polymorphism and atrial fibrillation among Han and Kazak residents of the Xinjiang region

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Abstract: Objectives: Atrial fibrillation (AF) is one of the most common and complex types of clinical arrhythmia syndromes. In recent years, an association between CYP11B2 gene polymorphisms and atrial myocardial fibrosis has received a significant amount of attention. This study explores the relationship between CYP11B2 gene -344C/T polymorphism and AF among Kazak and Han residents in the Xinjiang region and further clarifies the molecular mechanisms of atrial fibrillation. Methods: The study is a case-control study using traditional methods. We selected 156 Kazak and 203 Han patient cases in the Xinjiang region who had non-valvular atrial fibrillation as well as 307 Kazak and 418 Han cases of non-AF patients as a control group. Blood samples were collected, and DNA was extracted from the peripheral blood samples. The presence of the CYP11B2 gene -344C/T polymorphism was determined using polymerase chain reaction-restriction enzyme fragment length polymorphism (PCR-RFLP). Differences in the genotypes and allele distributions among the 2 groups were compared using Statistical Package for Social Science (SPSS) 17.0 statistical software. Student’s t test, the chi-squared test and logistic regression methods were used for the data analysis. Results: The genotypes of both ethnic groups followed a Hardy-Weinberg genetic equilibrium distribution. The 2 patient groups, compared with their respective control groups, showed significant dominant models in CYP11B2 gene -344C/T polymorphism genotype frequency and B1 allele frequency (P<0.05). The frequencies of the CYP11B2 gene -344C/T polymorphism in the Kazak patient group were higher compared with the control groups (P<0.05). The frequencies of the CYP11B2 gene -344C/T polymorphisms in the Han patient group was also higher compared with the control group (P<0.05). Logistic regression analysis showed that the frequencies of the CYP11B2 gene -344C/T genotypes were significantly different between the Kazak and Han patient groups and the control groups. Conclusion: CYP11B2 gene -344C/T polymorphism is associated with AF.

Keywords: Atrial fibrillation, CYP11B2 gene polymorphism, aldosterone

Introduction

Atrial fibrillation (AF) is the most common type of clinical arrhythmia. It increases the incidence and mortality due to stroke and sudden death. Aldosterone is a mineralocorticoid secreted by cells in the adrenal mineralocorticoid zone. It plays important regulatory roles in water-salt metabolism and sodium-potassium balance. Therefore, it is closely related to the occurrence and development of cardiovascular diseases [1, 2]. It has been found that multiple human tissues and organs, such as the kidneys, brain, vascular smooth muscle and so on, can produce aldosterone. The role of aldosterone in the cardiovascular system has received extensive investigation [3]. The renin-angiotensin-aldosterone system (RAAS) is closely related to the occurrence and development of various arrhythmias, such as atrial fibrillation. Aldosterone synthase (CYP11B2) is a key enzyme for the synthesis of aldosterone and is therefore an important component of the RAAS. This study investigated the relationship between the CYP11B2
CYP11B2 polymorphism and AF

gene -344C/T polymorphism and atrial fibrillation among Kazak and Han populations in the Xinjiang region.

Materials and methods

We collected blood samples from all of the participating subjects in compliance with the Declaration of Helsinki and under the ethical review of the Xinjiang Medical Association, The First Affiliated Hospital Ethics Review Board. All of the subjects provided written consent for blood collection.

The study subjects

We recruited 102 Kazak patients and 203 Han patients who were hospitalized in the Xinjiang Medical University Heart Center between January 2010 and June 2013 and diagnosed with atrial fibrillation. We also recruited another 54 Kazak patients from an epidemiological study of Kazak patients with atrial fibrillation in the Xinjiang region between 2008 and 2011 [4, 5]. This epidemiological study investigated 30,000 Kazak participants in the three northern regions of Xinjiang to determine the prevalence of atrial fibrillation among the Kazak people. The criteria to include patients with atrial fibrillation were as follows: patients 18-75 years of age and diagnosed with atrial fibrillation for over 6 months by 12-lead electrocardiogram (ECG) or 24-hour Holter monitor. Atrial fibrillation’s representation on ECG could include the following: 1) the disappearance of P waves and replacement by small irregular baseline fluctuations with irregular shape and amplitude known as f-waves; a frequency of approximately 350 to 600 beats/min; or an extremely irregular ventricular heart rate. The normal ventricular heart rate of atrial fibrillation patients who do not receive drug treatment and have normal AV conduction is usually between 100 and 160 beats/minute. The QRS complexes are usually normal. When the ventricular rate is too fast, differential conduction occurs, resulting in widening of the QRS complexes [6]. The criteria to exclude patients were valvular heart disease, congenital heart disease, hyperthyroidism, malignant tumor, acute inflammatory condition within the past 2 weeks and a family history of interracial marriage [7].

The control groups included 418 Han and Kazak 307 individuals who were patients undergoing medical examination in the First Affiliated Hospital of Xinjiang Medical University or patients hospitalized in the Heart Center between 2010 and 2012. There were no data about atrial fibrillation history or ECGs for the control patients. The diagnostic criteria for hypertension are based on multiple measurements of blood pressure on different days and include a systolic blood pressure ≥140 mmHg and (or) a diastolic ≥90 mmHg [8]. Drinking history was defined as drinking within the past 6 months. A history of smoking was defined by regular smoking within the past 6 months [9]. The diagnostic criteria for diabetes were as follows: 1. diabetic symptoms coupled with any glucose ≥11.1 mmol/L; 2. a fasting glucose ≥7.0 mmol/L; 3. a glucose ≥11.1 mmol/L after a 2-hour oral glucose tolerance test (OGTT). The OGTT result was based on World Health Organization requirements. Patients who met one of the above criteria underwent examination on another day. If they still met one of the three criteria, they were diagnosed as diabetic (1997 American Diabetes Association’s diagnostic criteria for diabetes).

Demographic characteristics for all subjects, including CYP11B2 genotypes, were unknown before data analysis.

Methods

Blood biochemical tests: Serum total cholesterol (TC), glucose, high-density lipoprotein (HDL), and low-density lipoprotein (LDL) levels were determined in the Laboratory of First Affiliated Hospital of Xinjiang Medical University.

DNA extraction: DNA samples were extracted from fasting venous blood collected on the second morning following patient admission. Venous blood was drawn into anticoagulant tubes containing ethylenediaminetetraacetic acid (EDTA) and immediately stored in a -20°C freezer. Genomic DNA was extracted using a whole blood genome extraction kit (Tiangen Biotech Co. Ltd., Beijing, China).

Primer design and PCR amplification of the CYP11B2 gene: The primers for the CYP11B2 -344C/T polymorphism are shown in Table 1.

The 20-µl polymerase chain reaction (PCR) system included the following: master mix, 10 µl; forward primer, 0.5 µl; reverse primer, 0.5 µl; double-distilled deionized water, 7.5 µl; and
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Table 1. Primers sequence of CYP11B2 -344C/T

<table>
<thead>
<tr>
<th>SNP</th>
<th>Primers sequence</th>
<th>Annealing temperature</th>
<th>PCR amplification products</th>
<th>endonuclease</th>
</tr>
</thead>
<tbody>
<tr>
<td>Rs1799998</td>
<td>Sense: 5’CAGGAGGAGACCCCATGTGAC3’</td>
<td>66.7°C</td>
<td>538 bp</td>
<td>Hae III</td>
</tr>
<tr>
<td></td>
<td>Antisense: 5’CCTCCACCTGTCCAGCCC3’</td>
<td></td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

Figure 1. Gel electrophoresis images of PCR amplification products: “M” represents Marker. There are 6 bands between 100 bp and 600 bp. “1-10” represents the 10 samples. Samples 1-10 had a 538-bp CYP11B2 gene PCR amplification product.

The completely mutated TT genotype showed three bands after enzyme digestion: 126 bp, 138 bp and 274 bp. The wild-type CC genotype showed four bands: 71 bp, 126 bp, 138 bp and 203 bp. The heterozygous genotype CT phenotype showed five bands: 71 bp, 126 bp, 138 bp, 203 bp and 274 bp. An image of the digested CYP11B2 -344C/T PCR product gel electrophoresis is shown in Figure 2.

Sequencing verification of CYP11B2 -344C/T: According to the band patterns of the CYP11B2 -344C/T samples, 20 specimens of each of the 3 genotypes were sequenced to verify the accuracy of the digestion gel electrophoresis results. The sequencing results are shown in Figures 3 and 4.

Statistical methods

The Statistical Package for Social Science (SPSS) 17.0 software was used for statistical analysis. Quantitative data were expressed as the mean ± standard deviation. Differences between groups were compared with t-tests. The genetic equilibrium was tested using the Hardy-Weinberg equilibrium. Counting and the Hardy-Weinberg equilibrium test were performed with the χ² test. In the logistic regression analysis, odds ratios and 95% confidence intervals (CIs) were used to describe the distribution of the major risk factors. P<0.05 was considered statistically significant.

Results

Comparison of the clinical data

Table 2 shows the basic clinical features of the Han and Kazak groups. There were no statistically significant differences in age or gender between the Han and Kazak case groups and control groups (P>0.05). Therefore, this study can be considered an age-matched and sex-matched case-control study. In the Han population, the AF group and the control group were significantly different with respect to rates of
CYP11B2 polymorphism and AF

Figure 2. Gel electrophoresis images of restriction-digested PCR amplification products: "M" represents Marker. There are 9 bands between 50 bp and 600 bp. Sample 1 is an empty well (no DNA sample); samples 3, 4, 7, 8, and 10 show the fully mutated TT genotype; sample 9 shows the wild-type CC genotype; samples 2, 5, and 6 show the partial mutant CT genotype. The 126-bp band and the 138-bp band have a similar size. The bands overlap and appear as a relatively wider band.

Table 3 shows the CYP11B2 -344C/T polymorphism genotype and allele frequency distributions. We conducted genetic equilibrium testing of the allele frequency distributions at this single nucleotide polymorphism (SNP) locus in the Han and Kazak populations. The results were in line with a Hardy-Weinberg equilibrium (P>0.05 between the atrial fibrillation group and the control group in both the Han and Kazak populations). In the Han population, the -344C/T locus genotype, co-dominant model and gene frequencies showed statistically significant differences (P<0.05) between the two groups. In the AF group, the TT genotype frequency (51.2% vs. 31.1%, P<0.001), CC + TT genotype frequency (55.7% vs. 46.7%, P=0.001) and T gene frequency (73.4% vs. 57.8%, P<0.001) were significantly higher than the control group. In the Kazak population, the dominant model and recessive model were significantly different. In the AF group, the CT + TT genotype frequency (67.9% vs. 62.2%, P<0.001) and TT genotype frequency (12.8% vs. 10.7%, P<0.001) were significantly higher than the control group. Logistic regression analysis suggested that CYP11B2 genetic polymorphism increased 1.874-fold risk for AF (Table 4).

Discussion

The aldosterone synthase gene is located on chromosome 8 (8q22); it is 7 kb long and includes 8 introns and 9 exons. The aldosterone synthase gene includes two genes: CYP11B1 and CYP11B2. Recent studies have shown that CYP11B2 catalyzes the final step of the in vivo biochemical synthesis of aldosterone. CYP11B2 is regulated by potassium and angiotensin II and is closely linked to cardiovascular diseases. CYP11B2 mutation frequently occurs at position-344, mainly as a transition from cytosine (C) to thymine (T). Another mutation frequently occurs within an intron. These mutations directly affect the transcription of CYP11B2, thus changing the gene’s expression level [10]. A number of studies have suggested
that the aldosterone synthase CYP11B2 gene-344C/T polymorphism is related to hypertension and left ventricular structure and function. Chronic high blood pressure results in compensatory adaptation, prompting atrial arrhythmia. Hypertension is the most common but treatable independent risk factor of atrial fibrillation (AF) [11, 12].

CYP11B2 gene polymorphism is closely related to plasma aldosterone levels, hypertension, atherosclerosis, cerebrovascular disease, left ventricular hypertrophy, myocardial infarction and coronary artery disease [13, 14], but some clinical studies have failed to support these views [15]. The evidence is also inconsistent for the association between CYP11B2 gene poly-

Table 2. Characteristics of included participants

<table>
<thead>
<tr>
<th></th>
<th>Han</th>
<th>Kazak</th>
<th>χ² or t</th>
<th>P value</th>
<th>Han</th>
<th>Kazak</th>
<th>χ² or t</th>
<th>P value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Age, Mean (SD)</td>
<td>53.17 (12.67)</td>
<td>54.85 (12.79)</td>
<td>1.552</td>
<td>0.121</td>
<td>53.85 (11.87)</td>
<td>51.87 (12.25)</td>
<td>-1.680</td>
<td>0.094</td>
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<td>Male/Female (%)</td>
<td>249/169</td>
<td>129/74</td>
<td>0.908</td>
<td>0.341</td>
<td>203/104</td>
<td>102/54</td>
<td>0.225</td>
<td>0.634</td>
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<tr>
<td>Hypertension (%)</td>
<td>75 (17.9)</td>
<td>80 (39.4)</td>
<td>33.617</td>
<td>&lt;0.001*</td>
<td>118 (71.0)</td>
<td>78 (50.0)</td>
<td>19.801</td>
<td>&lt;0.752</td>
</tr>
<tr>
<td>BMI, Mean (SD)</td>
<td>24.39 (3.97)</td>
<td>24.42 (5.35)</td>
<td>0.134</td>
<td>0.894</td>
<td>29.19 (4.46)</td>
<td>26.20 (4.67)</td>
<td>5.691</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>SBP, Mean (SD)</td>
<td>123.99 (16.45)</td>
<td>122.41 (22.68)</td>
<td>-0.885</td>
<td>0.377</td>
<td>127.85 (16.57)</td>
<td>139.04 (20.43)</td>
<td>5.925</td>
<td>&lt;0.001</td>
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<td>DBP, Mean (SD)</td>
<td>78.26 (11.84)</td>
<td>76.00 (12.23)</td>
<td>0.030*</td>
<td>0.772</td>
<td>85.53 (11.77)</td>
<td>7.072</td>
<td>&lt;0.001</td>
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Table 3. Frequency distribution of CYP11B2 -344C/T genotype and allele

<table>
<thead>
<tr>
<th>Rs1799998</th>
<th>Han</th>
<th>Kazak</th>
<th>χ² or t</th>
<th>P value</th>
<th>Han</th>
<th>Kazak</th>
<th>χ² or t</th>
<th>P value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Genotype</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>CC</td>
<td>65 (15.6)</td>
<td>116 (37.8)</td>
<td>9 (4.4)</td>
<td>50 (32.1)</td>
<td>130 (31.1)</td>
<td>33 (10.7)</td>
<td>20 (12.8)</td>
<td>0.449</td>
</tr>
<tr>
<td>CT</td>
<td>223 (53.3)</td>
<td>158 (51.5)</td>
<td>90 (44.3)</td>
<td>86 (55.1)</td>
<td>104 (51.2)</td>
<td>3 (1.0)</td>
<td>353 (84.4)</td>
<td>0.133</td>
</tr>
<tr>
<td>TT</td>
<td></td>
<td></td>
<td></td>
<td></td>
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<tr>
<td>Dominant model</td>
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<tr>
<td>CC</td>
<td>65 (15.6)</td>
<td>116 (37.8)</td>
<td>9 (4.4)</td>
<td>50 (32.1)</td>
<td>130 (31.1)</td>
<td>33 (10.7)</td>
<td>20 (12.8)</td>
<td>0.449</td>
</tr>
<tr>
<td>CT + TT</td>
<td>353 (84.4)</td>
<td>191 (62.2)</td>
<td>194 (95.6)</td>
<td>106 (67.9)</td>
<td>149 (48.5)</td>
<td>274 (89.3)</td>
<td>138 (87.2)</td>
<td>&lt;0.001*</td>
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<tr>
<td>Recessive model</td>
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<tr>
<td>TT</td>
<td>130 (31.1)</td>
<td>33 (10.7)</td>
<td>149 (48.5)</td>
<td>106 (67.9)</td>
<td>138 (87.2)</td>
<td>136 (87.2)</td>
<td>0.259</td>
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<tr>
<td>CT + CC</td>
<td>288 (68.9)</td>
<td>138 (87.2)</td>
<td>99 (48.8)</td>
<td>136 (87.2)</td>
<td>138 (87.2)</td>
<td>136 (87.2)</td>
<td>0.259</td>
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<tr>
<td>Co-additive model</td>
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<tr>
<td>CT</td>
<td>223 (53.3)</td>
<td>158 (51.5)</td>
<td>90 (44.3)</td>
<td>86 (55.1)</td>
<td>104 (51.2)</td>
<td>33 (10.7)</td>
<td>20 (12.8)</td>
<td>0.449</td>
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<tr>
<td>CC + TT</td>
<td>195 (46.7)</td>
<td>149 (48.5)</td>
<td>137 (55.7)</td>
<td>70 (48.5)</td>
<td>137 (55.7)</td>
<td>137 (55.7)</td>
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<tr>
<td>Allele frequency</td>
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</tr>
<tr>
<td>C</td>
<td>353 (42.2)</td>
<td>390 (63.5)</td>
<td>108 (26.6)</td>
<td>186 (59.6)</td>
<td>298 (73.4)</td>
<td>224 (36.5)</td>
<td>126 (40.4)</td>
<td>0.247</td>
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<td>T</td>
<td>483 (57.8)</td>
<td>224 (36.5)</td>
<td>298 (73.4)</td>
<td>126 (40.4)</td>
<td>298 (73.4)</td>
<td>224 (36.5)</td>
<td>126 (40.4)</td>
<td>0.247</td>
</tr>
</tbody>
</table>

Note: *indicated significant difference compared to control.
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morphism and hypertension, atherosclerosis and other diseases in patients of different ethnic groups and in different regions. These studies suggest that CYP11B2 gene polymorphisms could be different among people of different ethnic groups and regions [16]. One study found that aldosterone level is closely linked to the CYP11B2 -344C/T polymorphism. Individuals with the TT and CT genotypes have significantly higher levels of aldosterone than those who have the CC genotype [17].

Clinical studies have shown that aldosterone antagonists may help reduce the incidence and persistence of AF. Angiotensin II receptor antagonists have been administered to AF patients [18]. However, the association between the CYP11B2 -344C/T polymorphism and AF has rarely been studied. Studies have confirmed that the occurrence and development of AF is genetic. The CYP11B2 gene may have polymorphisms at other sites that can affect the levels of plasma aldosterone and angiotensin II or that may induce atrial fibrillation by other mechanisms. Aldosterone induces increased blood volume, blood pressure, left ventricular pressure, left ventricular hypertrophy, left atrial pressure, and left atrial volume. In addition to the indirect effects of aldosterone on cardiovascular function, aldosterone can also have a direct effect on the heart. In animal experiments, scientists found that aldosterone could directly induce cardiac hypertrophy and fibrosis [19-21]. The binding of aldosterone to its receptor promotes the proliferation and differentiation of myocardial cells and vascular smooth muscle cells. Aldosterone signaling also promotes fibroblast proliferation and hypertrophy, leading to a significant increase in collagen production. These effects lead to cardiac fibrosis and cardiac structural remodeling, and can trigger heart rhythm disorders. In addition, the binding of aldosterone to mineralocorticoid receptors can activate the RAAS and strongly induces myocardial fibrosis. In summary, aldosterone may directly or indirectly cause atrial enlargement and atrial fibrosis, leading to structural and electrical atrial remodeling. These effects may induce and maintain atrial fibrillation. Therefore, the CYP11B2 gene-344C/T polymorphism may be related to atrial fibrillation, or AF may be due to other ethnic, environmental and genetic factors.

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Disclosure of conflict of interest

None.

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